

Marinho Del Santo¹

Reconstitution of the immunological defence and *Candida albicans* infection in oral mucosa of HIV+ patients under HAART

ABSTRACT:

The HIV infection is a worldwide spread disease which with the HAART (highly active antiretroviral therapy) application has become a chronic disease. The HAART promotes the reduction of the HIV viral load and partial and temporary reconstitution of the immunological defence system of the HIV-infected subject, although for that its toxicity and patient adherence to the treatment might be well monitored. With the HAART, the past high prevalence of oral and oropharyngeal lesions decreased significantly, although in a non-homogeneous pattern. The fungus *Candida albicans* is a commensal microorganism of the human gut tract which provokes an opportunistic infection, when there is an imbalance between its virulence and the defence conditions of the host. The pathogenicity of the *Candida albicans* influences the degree of opportunistic infection; however, the fungal colonization is mainly dependent of the current immunological status of the patient. The host defence against *Candida albicans* is also provided by non-immunological barriers, physical as the keratinocytes of the oral epithelium, serological as the neutrophils, polymorphonuclear leukocytes and macrophages or humoral as the saliva, although the role of the salivary immunoglobulins is still unclear. Independently of the immunosuppression, the sensitive control to balance immunological innate and immunological acquired actions is complex and it prevents against an indiscriminate immunological acquired response. Dendritic cells and lymphocytes are the main defensive immunological cells of the oral mucosa. The dendritic cells phagocytise and deplete microorganisms, presenting the products of such depletion as antigens to the T lymphocytes, which provide acquired immunological defence for excellence. Specific Th1 type provides cell-mediated immunological protection against *Candida albicans* and other pathogens. Moreover, Th2 type cells provide immunological tolerance against external and auto-antigens. Treg and Th17 cells are actors of vital importance in the switching between Th1 type and Th2 type responses, although the complete understanding of their roles in this balance is still an ongoing process.

Keywords: candida albicans; HAART; HIV+ patients; immunological defence; oral mucosa.

¹ DDS, PhD. Orthodontist in Private Practice and Private Lecturer.

Declaration of Interests: The authors certify that they have no commercial or associative interest that represents a conflict of interest in connection with the manuscript.

Corresponding Author:

Marinho Del Santo.
E-mail: marinho@delsanto.com.br

DOI: 10.18762/2525-5711.20160005

Submitted:
Accepted:
Originally published as 1 (2) April/June 2012.

INTRODUCTION

The HIV infection is a devastating epidemic, with serious socio-economical and population reduction consequences. The main strain of the virus which causes acquired immunodeficiency syndrome is the HIV1, in this present paper called HIV. Few HIV+ patients do not present oral lesions during some phase of the disease progress, presenting pathological signals and symptoms mainly in oral mucosa and salivary glands¹³¹.

It is known that the mucosa of HIV+ patients are vulnerable to the incidence of opportunistic fungal infections, as candidiasis, cryptococcosis, invasive aspergillosis, disseminated histoplasmosis and disseminated coccidioidomycosis^{146,285}. However, the compromising of oral defence in HIV+ patients occurs even before the incidence of opportunistic infections. It has been mentioned, as examples, the occurrence of salivary glands dysfunctions²⁰¹ and the presence of yeasts and hyphae of *Candida albicans* in oral mucosa^{345,316} before significant decrease in the number of CD4+ cells in the blood stream and in the IgA concentration of the saliva of infected patients³¹⁶.

Therefore, the current research in the oral aspects of HIV+ patients highlights a broader approach, involving not only the classical deficiency of CD4+ cells but especially the innate and acquired attributes of the immunological system of such subjects. Researchers investigate in special the defensive role of cells Th1 and Th2 and the regulatory role of cells Treg and Th17, contextualized in the complex control of the interactions between microorganisms and host.

In such contextualized approach, it is mandatory to consider the proofed partial and temporary reconstitution of the immunological system of HIV+ patients under HAART (highly active antiretroviral therapy), with important reduction of the prevalence of opportunist infection by *Candida albicans* in oral mucosa.

HAART: HIGHLY ACTIVE ANTIRETROVIRAL THERAPY

With HAART application, the immunological system of the HIV patient is partially and temporarily reconstituted²⁷, due to the decrease of the viral load and the enhanced response of the defensive cells, in adults^{19,83,9,332} and children³³⁵. More than the increase in number of the defensive cells, with increase of the hematopoietic activity³³², there is a functional recovery of immunodepressed cells^{232,332}.

Under HAART, the enhanced reactivity of cells CD4⁺^{19,20,83,232,89,90}, and other effects imply in significant decrease of the morbidity and mortality of HIV+ patients^{231,74}. The positive effects of HAART may be also of non-immunological nature⁵⁰, as a worst fungal adherence to epithelial cells²⁸.

Moreover, it has been suggested that HAART could act directly as anti-fungal drug, especially over the virulence factor Sap (secretory aspartyl proteinase) of *Candida albicans*^{48,49,216}. Microbiologically, certain fungal strains may respond differently to HAART. It was verified an increase of *Candida spp. (no-albicans)* in the oral microbiota, as examples, *C. tropicalis* e *C. parapsilosis*; however, there were rare assessments of *C. dubliniensis*, *C. norvegensis*, *C. humicola* and *C. rugosa*²⁰⁷.

Classically, HAART includes at least two inhibitor drugs of reverse transcriptase of nucleoside (RTI) plus a protease inhibitor (PI) or a reverse transcriptase inhibitor of no-nucleoside²²⁶. In regard to the risk-benefit of its composition, toxicity is the main cause to avoid certain regimens; therefore the clinical protocols must always evaluate the side effects simultaneous to the viral load reduction⁸⁸.

In theory, reduction of the antiretroviral drugs included in the medication could be worth, since it implies in lower toxicity and better adherence of the patient to the treatment. However, simplified regimens carry a significant higher risk of resistance and consequent loss of the power of viral suppression²⁷¹. In HIV+ patients the resistance to HAART may be due to the high rate of viral mutations, since the applied drugs have selective effects^{226,241}.

In opposite, very "efficient" medications, there is a high risk of substantial side effects, what may provoke important lack of commitment of the patient with the treatment; what also significantly diminish its efficacy^{320,325}. As consensual rule, the therapeutic prescriptions, if working well, must be preserved, unless a change is clinically necessary²⁷¹.

The IRIS (Immunologic Reconstitution Inflammatory Syndrome) occurs days or weeks after the beginning of the antiretroviral therapy, as an organic response to the drugs which compose HAART²⁷. The hypertrophy of the parotid gland is suggested as a possible oral manifestation of IRIS in patients under HAART²²⁹.

With HAART, the HIV infection has become a chronicle disease and it presents a different scenario. One of the features of this new scenario is the expressive reduction of the incidence and prevalence of opportunistic oral lesions^{19,83,9,199,286,20,15,172,278,86,317}.

Epidemiologically, studies confirm such reduction as definitive trend in the USA^{160,142}, in Mexico²⁵², in general industrialized countries¹⁴², and in Brazil^{109,137,243}. However, in England there is no evidence that it has happen a significant difference in the reduction of the incidence of oral candidiasis with the HAART introduction, substituting no-HART (antiretroviral therapy not highly active), according to Ives et al¹⁵².

It is important to highlight that cohorts which follow patients submitted to HAART for a long period of time, the immunological reconstitution might be much more expressive³³⁴,

although such recovery is slow and incomplete in immunological very compromised patients^{209,310}. In these patients, the re-incidence of oral lesions, especially candidiasis, may be an indicator of HAART failure^{142,253,211}.

Besides that, the reduction of the incidence of oral manifestations in HIV+ patients was not homogeneous for all lesions²³⁵, being hard to distinguish the oral manifestations of the HIV infection from the side effects of HAART¹⁴². As an example, HAART significantly increased the presence of oral warts^{129,142,109}.

Other factors must also be considered in this new scenario. Co-infections may in smaller decrease of incidence of oral lesions¹²² and socio-demographic factors may also influence, since there is a trend for the decrease to be smaller in HIV+ patients under HAART Who present a lower scholar level⁷⁵.

FUNGUS CANDIDA ALBICANS

Candida albicans is an imperfect diploid dimorphic fungus, with phenotypic flexibility, which resides in a commensal way in the human gut in 40% of healthy subjects, usually without potential to overwhelm the host immunological defence¹⁴. The possibility of *Candida albicans* colonize, penetrate and damage the host tissues depend basically of the unbalance between the fungal virulence and the defence conditions of the host, immunological^{148,111,112,84} or no-immunological, as the pH of the anatomic site colonized by *Candida albicans*⁷⁶ and the possibility of formation of a shelter niche, a biofilm for homing and resistance of the microorganisms⁵⁹. In this case, the fungus *Candida albicans* does not present the commensal status anymore and provokes an opportunistic infection called candidiasis.

Candidiasis can be a light opportunist infection or up to a live-threatening disease in seriously immunodepressed patients. To achieve such level of severity, the fungal infection, of species *C. albicans* and *no-albicans*, occurs through the mucosa and gain the blood stream leading to a generalized systemic candidiasis⁸². That is a potentially lethal complication in AIDS patients in advanced stage of the disease^{171,315,240}.

The pathogenicity of the fungus *Candida albicans* is complex and multifactorial. The secretion of hydrolytic enzymes aspartyl proteinases (Saps) promotes a virulence potential well described in the pertinent literature^{70,149,78}. The enzymes Saps are codified by at least ten genes Sap (Sap1 a Sap10), identified by mRNA sequences^{148,264}, which roles in the colonization and invasion of the host tissues are distinct^{148,213}. The phenotype of the opportunistic fungus *Candida albicans* influences the cytokines production and the response of the host to the infection³²⁸. First, a fungal aggression stimulates an innate response, constituted by phagocytosis, generation of pro-inflammatory mediators,

traffic of inflammatory cells to the injury site and the beginning of an acquired immunological response¹⁴⁵.

The pattern of adherence and fungal colonization of the epithelial cells of the oral mucosa reflects its pathogenicity, mainly because of the expression of Saps by different strains and biotypes of *Candida*³⁹. In HIV+ patients, the higher adherence of *Candida albicans* to the oral mucosa, independently of lower levels of antibodies against *Candida* in saliva and of potential lower salivary secretion, is simultaneous with the Sap production³⁰⁵. Such results suggest that there is a selective colonization of *Candida albicans* strains which present better adherence to the oral mucosa^{228,76,343}. Other enzyme secreted by *Candida albicans* which plays a pathogenic role is the B phospholipase^{151,123}, that can kill or damage the host cells¹²³.

Ultra-structural studies showed that the tissue response in oral mucosa of HIV+ patients is different in the pseudomembranous candidiasis when compared to the erythematous candidiasis. In the pseudomembranous form, the cellular immunological compromising, especially of dendritic cells and lymphocytes, is proportionally more severe than in the erythematous form²⁶⁸. In the pseudomembranous form, the fungal hyphae are abundant and extend up to the spinous layer (*stratum spinosum*) of the oral epithelium, with simultaneous parakeratosis, acanthosis and spongiosis of the infected epithelium. The hyphae penetrate in inter-cellular spaces, suggesting that *Candida albicans* may present thigmotropism (guide by contact), observed in vegetal fungi and recognized in fungal proliferation *in vitro*²⁸⁷. The inter-cellular fungal penetration is facilitated by the detachment of epithelial desmosomes, probably caused by Saps and/or phospholipases produced by *Candida*²⁵⁹, also observed in HIV- patients²¹². It is interesting to observe that in the HIV+ patient the immunological cellular reaction against fungal hyphae seems to be minimal; although, possibly, because of the ongoing immunodepression²⁵⁹. In the erythematous form, hyphae are rare^{259,104,258}.

There is important variation of strains of *Candida albicans* and other species, which colonize the oral cavity of HIV+ patients⁸⁷. The great majority of HIV+ patients who present oral candidiasis are mainly infected by the endogenous *Candida albicans*, already present as a commensal microorganism of the oral flora of the patient. However, part of the patients presents new *Candida albicans* strains^{275,189,23} or other *Candida no-albicans* species^{170,326}, as *Candida dubliniensis*^{64,303,206,280} and *Candida glabrata*¹¹⁴. Different strains and species may be transmitted between subjects²⁷⁵, what may contribute for episodes of fungal resistance to therapeutic drugs.

Overall, there is significant genetic diversity⁷² and the degree of fungal colonization increases proportionally to the disease advancement^{246,326}, depending directly upon of the

individual response in each anatomical infected site^{289,113,115,111,112}. Therefore, occurrence of oropharyngeal candidiasis and vaginal candidiasis are not associated⁷².

The individual resistance to drugs as fluconazole may occur because of the use in different episodes of candidiasis or prolonged use^{193,110}, and contributes for the diversity of strains and species presented by the patients^{275,255,23,170}. Moreover, interactions between the HIV virus and *Candida albicans* may change the virulence potential of the fungus¹³⁶.

ORAL INFECTION BY *CANDIDA ALBICANS* IN HIV+ PATIENT

The earliest and significant incidence of oral and oropharyngeal lesions in HIV+ patients was presented in the 1980's^{126,1,132,164} as a predictive signal of the HIV infection^{132,164,308,272,162,222,237,46,67}.

In 90% of the HIV infected individuals, in some stage of the disease, it occur one or more episodes of *Candida albicans* infection¹¹², which can affect either the oropharyngeal region as the oesophagus^{91,120}. In 75% of the cases of oropharyngeal candidiasis, it also occur esophageal candidiasis or significant risk of its occurrence³.

Esophageal candidiasis is only confirmed by endoscopic biopsy²⁵⁶, and part of the patients positive for that (30 to 43%) do not present symptoms, as pain and burning sensation¹¹⁶. If they do, they must receive the prescribed anti-fungal therapy even without diagnosis confirmation by endoscopic exam^{250,12}. With HAART, episodes of opportunistic oropharyngeal infections, sometimes called "AIDS predictors", decrease significantly⁴⁴.

The resistance of the oral mucosa to candidiasis in a health subject is the sum of the redundant mechanisms which include salivary anti-candidiasis proteins, inhibition of the growth of *Candida albicans* by oral keratinocytes and the acquired immunological response provided by T lymphocytes⁸⁴. The protection of the salivary proteins and the action of the oral keratinocytes against *Candida albicans* was evidenced *in vitro*. Experimental models of oropharyngeal candidiasis have detected that the mechanisms and the role of mediators in the acquired immunological response against *Candida albicans*, with presentation of antigens by the dendritic cells to CD4⁺ T lymphocytes⁸⁴. However, the presentation of antigens by keratinocytes is uncertain, since these cells are located in the superficial layer of the epithelium and the CD4⁺ cells are located in the basal layer²⁹², although such presentation may be stimulated by *Candida* infection¹⁶.

As a model to study the evolution of the incidence of candidiasis, it has been suggested that the debility and immaturity of the dendritic cells may interfere in the presentation of *Candida albicans* antigens to the CD4⁺ cells, which are debilitated by the HIV infection. HIV virus may also prejudice the phagocytary

activity in the oral mucosa against *Candida albicans*, leading to clinical infection. However, such debilities may be partially compensated by the defence mechanisms still preserved (physical barrier of the keratinocytes, citotoxic activity of the CD8⁺ lymphocytes and partial phagocytary activity). Such remaining mechanisms may limited the candidiasis proliferation in the oral mucosa and prevent its systemic dissemination⁸⁴.

Moreover, dendritic cells, T lymphocytes and macrophages of the oral mucosa may be the entrance door for the HIV viral infection⁵⁸, although the transmission of the HIV virus by oral mucosa is unexpected¹⁵⁰.

In general, oral lesions in HIV+ patients have been extensively categorized^{2,130,337,67} and directly correlated with the decrease of the CD4⁺ lymphocytes number^{108,270,200,124,163,166,181} and with the HIV viral load^{197,133}. Different opportunist infections are associated with the viral load, but not with the number of CD4⁺ cells⁴³, although the number of CD4⁺ cells is indicative of the stage of evolution of the HIV infection and the baseline for therapeutic decisions²⁹⁴.

Among the detected oral lesions, candidiasis is the one with greater prevalence and incidence, although the epidemiological data is very heterogeneous. There are some reasons for such heterogeneity: a) differences among the assessed samples and the stage of the HIV infection in the included research subjects; b) concomitant prevalence of other oral lesions, which may difficult the differential diagnosis of candidiasis¹²⁴; c) significant influence of covariants as smoking habit^{233,266}, use of alcohol²³⁶, use of heroin/methadone¹³³ and oral hygiene and; d) prevalence of co-infections potentially facilitators of the fungal colonization, as the *Herpes simplex* virus (HSV) and the Epstein-Barr-EBV virus²⁸⁸.

Oral candidiasis may be presented in the pseudomembranous form, erythematous form, angular cheilitis¹²⁸ and hyperplasic²⁵⁸. The pseudomembranous and erythematous forms are the most common²⁶⁰. The pseudomembranous form is characterized by the presence of white papular multifocal lesions. The diagnosis is mainly clinical but the diagnosis confirmation is made by microbiological culture of clinical collection, what leaves a reddish surface. Fungal hyphae are pathognomonic. It practically does not present associated inflammation and rarely presents micro-abscess, even though the colonization area is broad^{104,258}.

The erythematous form provokes multiple micro-abscesses in the epithelium^{260,104} and diffused erythema in the palate, oropharynge and tongue dorsum. In general, fungal hyphae are absent. The erythematous form demands biopsy for diagnosis confirmation. In the hyperplasic form, a superficial cellular reaction occurs against the pathogen, depending upon the degree of its virulence²⁵⁸.

Signs of oropharyngeal fungal infection vary from light to generalized thrush³²⁶. The esophageal candidiasis can also be light or generalized, depending upon the stage of AIDS, or can be associated with an acute HIV infection^{55,239}. The patient may present hyperplastic palatal papillae²⁶⁰ or exfoliated cheilitis, mainly in the lower lip²⁶¹.

In regard to the symptoms, the patient with oropharyngeal candidiasis presents burning feeling, pain, taste change and difficult to swallow liquid and solid food¹²⁸. Esophageal candidiasis may lead to dysphagia, odynophagia, fever and nausea/vomiting^{308,60}. Because of painful swallowing, the limited intake of food and liquid may provoke expressive weight loss, which is very common in HIV+ patients³⁰⁸.

SALIVA

The salivary flow and its aggregating properties provide a dynamic balance between *Candida albicans* and other commensal microorganisms of the oral microbiota, protecting against the establishment of oral candidiasis in a healthy subject^{26,196,140}. However, such salivary mucine properties may also facilitate the *Candida albicans* adherence to the oral mucosa^{101,143}.

Some salivary proteins present fungicidal effects. Lysozyme and lactoferrin are two proteins of the innate defence, no-immunological and no-specific against *Candida albicans*; however, with potential fungicidal properties^{141,290,223,344,274,139}.

Histatins are other salivary proteins which may contribute to the non-immunological innate defence of the oral mucosa^{254,100,314,139}, as the antileukoprotease³¹².

In HIV+ patients, the salivary antifungicidal effect is controversial. It is lower for a group of researchers¹⁸³; however, for others, the salivary lysozyme concentration is greater^{345,18,195} and the lactoferrin production is not definitively associated to the limited proliferation of *Candida albicans*⁸⁴.

Candidiasis and salivary flow may also be associated. Subjects with Sjögren syndrome present reduced salivary flow and higher incidence of candidiasis.²⁶³ The same occurs with HIV+ patients in advanced stage, in which the salivary flow is reduced in 40%¹⁸³ and in patients with oral acid pH, in which the virulence of *Candida albicans* is enhanced^{273,176,77}.

The detection of specific IgA antibodies against *Candida albicans* suggests that there is a specific humoral response against *Candida albicans* that inhibits the adherence and colonization of such fungus in the oral epithelium; however, such hypothesis was confirmed *in vitro* only^{103,330} and the fact that subjects with deficient salivary IgA production do not present significant increase in the incidence of candidiasis⁷ makes such hypothesis vulnerable.

HIV infection produces direct and indirect effects in the humoral and cellular immunity of the oral mucosa, innate or acquired⁵⁷, with consequent increase in the incidence of

opportunistic infections; however, conclusions in regard to the humoral immunity of HIV+ patients, especially about salivary flow and salivary IgA concentration, are controversial^{56,111}.

For some authors, there is no significant alteration in the salivary flow, although there is a tendency for flow reduction¹⁹⁵. For others, the reduction is certain and consequently its antimicrobial effect too¹⁸³. According to some authors, it occurs significant reduction in the IgA production and consequent reduction in the antimicrobial effect^{215,306}. However, for others, there is no change²⁹ or the IgA anti-*Candida* production increases^{18,66}, simultaneously with the increase of the production of anti-microorganism proteins as lactoferrin, lysozyme and histidine; independently of the decrease in the salivary flow^{68,99}.

The change in the profile of the immunological response from Th1 to Th2 might be critical in the immunological imbalance in HIV+ patients⁶². Healthy subjects present in the saliva cytokines of Th1 and Th2 immunological responses. However, in HIV+ patients, the profile of salivary cytokines is clearly of Th2 response but not Th1 response¹⁷⁵.

IMMUNOLOGICAL DEFENCE

Immunologically, the host defence can be divided in innate and acquired. The innate defence is congenital and DNA oriented and the acquired defence is basically organized by T and B lymphocytes with structurally unique receptors. The lymphocyte receptors are random generated, and provide an extremely diverse repertoire of defence. Then, there is a great probability that a lymphocyte recognizes an antigen and, consequently, to be activated and proliferate in cloned expansion. Such process is absolutely necessary for an efficient immunological response²⁰⁴. The effector mechanisms of the innate immunity, including macrophages, phagocytes and complement system, are immediately activated when an antigen is presented to the host, while the cloned expansion delays in average from 3 to 5 days²⁰⁵.

The activation of the acquired immunological system can be triggered not only by infectious microbial antigens, but also by environmental innocuous antigens and self-antigens, generating allergic and auto-immune diseases²⁰⁵. So, how the immunological system can identify the origin of the antigen? And when the immunological response must be activated? The connections among some components of the immunological system are not well understood yet, however, recent progresses allow a contextualized view of the defence system²⁰⁴ and its failure substantially collaborates for the susceptibility of the oral mucosa to candidiasis in HIV+ patients¹²⁵.

The innate immunity is fundamental in the host defence against pathogenic antigens. It is mediated by many genetically pre-determined receptors, which specificity is molded by natural selection. The issue is that the genome can codify only

a limited number of gens, for example, the human genome contains only 75,000 to 100,000 gens, which, in the most cases, are not related to the immunological recognition²⁰⁵. In opposite, the acquired defence system presents approximately 10^{14} receptors for immunoglobulins and 10^{18} lymphocytary receptors, developed in a clonal basis. With such defence *armamentarium*, even though the microorganisms being extremely heterogeneous and suffering periodic mutations, the acquired defence can potentially recognize ever possible antigen. However, the trade off of such diversity is the lack of ability to distinguish pathogenic external antigens from innocuous external antigens and from self-antigens.

The strategy of innate defence is not clonal as the acquired one and it does not recognize ever antigen *per se*, however, be triggered by few molecular standard structures present in large groups of pathogenic microorganisms, as example, bacterial lipopolysaccharides, peptidoglycans, lipoteichoic acids, mannans, bacterial DNA, double-stranded RNA and glucans¹⁵⁴. For example, lipopolysaccharide is synthesized only by bacteria, and the receptors for such molecules alert the host to the presence of an infection by bacteria. Such “sensitive” and sophisticated balance can prevent the invasion of pathogens and, at the same time, preserve the symbiotic interaction with the commensal flora^{21,276,97}.

However, other important effect of the innate immunologic defence is the professional antigen-presentation, especially by dendritic cells, macrophages and B lymphocytes. In general, when a molecular pattern in a pathogenic microbe is recognized, antigen-presenting cells (APCs) process it and present part of that, as example, MHC (major histocompatibility complex) class II segments.

In order to trigger the acquired immunological system, beside MHC class II presentation, co-stimulatory signals as CD80 and CD86 molecules are necessary. The induction of expression of such molecules is also controlled by the innate immunological system, throughout the activation of toll-like receptors (TLR) in an infectious scenario. The recognition of an antigen by a T cell in the absence of CD80 or CD86 molecules promotes its permanent inactivation. Then, the combined activation of different receptors, TLR or non-TLR, results in complementary effects, synergic or antagonic, which modulate the innate and acquired immunity^{313,97} and protect against an indiscriminate acquired immunological stimulation⁶³.

Systems of receptors may modulate the antigenic specificity of the response, as T helper 1 (Th1) or T helper 2 (Th2), throughout the feedback of the effect cells to the dendritic cells and not throughout the instructions provided by the pathogens, therefore, an experience-based criteria, inducing and maintaining an appropriated polarized response¹⁵⁹.

ORAL MUCOSA INVASION

Histologically, the oral mucosa presents in 60% of its surface similar characteristics to the esophagic and vaginal mucosa. The stratified squamous epithelium and the lamina propria of the connective tissue, mainly formed by dense collagen fibers, are separated by a basal membrane. One difference between the oral epithelium and the esophagus/vagina epithelium is the oral keratinized epithelium, which is similar to the skin epithelium, is found in the gingiva and hard palate and represents 25% of the oral mucosa. Other difference is the dorsal tongue epithelium, which presents a large number of sensorial gustative papillae, representing 15% of the oral mucosa surface.

Keratinocytes are cells of the oral epithelium adjacent to the basal membrane, which united by desmosomes (in larger number and better attached in the external region of the epithelium) provide the main physical barrier against pathogenic agents invasion. The oral epithelium turnover (approximately 14-20 days) occurs due to the lost of the protein integrin of the keratinocytes. Such process is fundamental for the homeostasis of the oral mucosa, limiting for example, the colonization and infection by *Candida albicans* fungus²⁷⁴.

Epithelial cells invade the lamina propria, allowing that dendritic cells present antigens to lymphoid tissue nodes, which contain lymphocytes as host defence agents. Keratinocytes are HIV infectable cells²⁴⁹, with potential risk that their action to be diminished, although such hypothesis has not been clinically proofed²⁹¹. The calprotection production in keratinocytes, preserved in HIV+ patients, is a physical barrier against the penetration of *Candida albicans* hyphae¹⁰⁴.

In the skin, infected keratinocytes by *Candida albicans* produce specific cytokines which collaborate to the immunological response regulation^{11,295,277}, as in the oral mucosa^{179,117,292,95,96,214,94}, thru the activation of innate recognition mechanisms by toll-like receptors (TLR). Furthermore, epithelial cells might secrete antimicrobial peptides as beta-defensins, which prevent the installation of the infectious process in the oral mucosa³³⁵.

Neutrophils offer innate protection, mainly phagocytizing and digesting bacteria and fungi, and also producing cytokines which attract and stimulate other immunological actions, instructing and modulating dendritic cells^{282,321}.

The local response against *Candida albicans* is mediated by macrophages and polymorphonuclear leukocytes, which are more potent than the dendritic cells to kill *Candida albicans*²²⁰ and play an important role in the innate immunological response²⁰⁵. Further, they stimulate the lymphocytary proliferation and the synthesis of related cytokines^{17,336,125}.

Macrophages are physiologically located in the lamina propria and produce peroxy-nitrite, an *anti-Candida* product³²⁷. They present a repertoire of receptors which promote the ho-

meostasis, defence and immunological induction^{309,198}. When they are activated by cytokines as interferon-gamma, they differentiate and participate of the acquired immunological response against *Candida*³²⁷.

Polymorphonuclear leukocytes are present in the blood stream, providing protection against systemic infections^{338,112}, and are also in the lamina propria and in the epithelium by inflammatory induction²⁰⁵.

In HIV+ persons, the phagocytary function of the macrophages is not affected^{1225,125}. Macrophages may also produce nitric acid, an *anti-Candida* product. Such production may be regulated by T gamma-delta cells¹⁵⁵, and is not compromised in HIV+ patient¹²⁵. However, cytokines as IL4 e IL10 may compromise the antifungal action of the polymorphonuclear leukocytes, increasing the susceptibility of the host to opportunist infections³⁰⁷.

Dendritic cells and lymphocytes are the main acquired immunologic cells *anti-Candida* of the oral epithelium²²¹. Dendritic cells phagocytose *Candida*, presenting the products as antigens to the T lymphocytes, which are the immune cells by excellence. The proliferation of specific lymphocytes against *Candida* is stimulated by cytokines produced by dendritic cells²²¹.

Dendritic Cells

Dendritic cell had its identity and function clarified in the 1970s^{297,298,300,296,299}. Langerhans cells are a sub-population of the dendritic cells^{22,251}, a type which presents certain features, as example, CD1a⁺ identification, Birbeck granules, Lag antigens and E-cadherin^{53,52}.

Dendritic cells are located in the basal and supra basal layers of the epithelium of the oral mucosa^{40,73,5,257,69,323,185,24,283,284,71,158}, architecturing the MALT (mucosal associated lymphoid tissue) as primary lymphatic tissue. In oral mucosa, the dendritic cells and other antigen-presenting cells must quickly respond against intrusion pathogens²²⁰. Similarly, in the gut the dendritic cells architecture the GALT (gut associated lymphoid tissue)¹⁸⁶; however, there they are considered secondary lymphatic tissue⁴⁰. Anyway, in both anatomic sites they are fundamental for the acquired immunological protection²⁴⁸. Furthermore, in the oral mucosa they might be more efficient in the antigen-presentation process to T lymphocytes than the skin dendritic cells¹³⁸.

Dendritic cells are specialized in the antigen capture, migration and presentation to T lymphocytes^{346,54,340,22,158}, performing a crucial defence against pathogens^{322,324}. Furthermore, the dendritic cells might collaborate to the immunological tolerance of the subject against self-antigens, minimizing auto-immune response²². In a broader view, dendritic cells also perform diverse roles in the mobilization of the immunological response, innate or acquired, working simultaneously in the homeostasis and host protection¹⁵³.

The functional properties of the dendritic cells are related to their state of maturation²³⁸. Different lineages and phenotypes of dendritic cells have being identified and there are signals that the Langerhans cells come from the same lineage of lymphocytes CD8⁺¹⁰. Mature dendritic cells induce T helper 1 (Th1) response and immature dendritic cells inhibit the proliferation of Th1 and induce T CD4⁺ regulatory cells (Treg) and the IL-10 production¹⁵⁶.

Treg cells stimulate the CTLA-4 production, which negatively regulate T citotoxic cells. Interferon-gamma, IL4 e IL12¹⁰² are required to induce CD4⁺ lymphocytes and Th response, possibly by combined innate and acquired immunological mechanisms^{208,98}. The IL18 cytokine has a similar action to the IL12 cytokine and stimulate Th1 response; however, it could also stimulate the tolerance response of the Th2 type, becoming an example that the immunological protection is heterogeneous and complicated²¹⁸.

The fundamental question in the ontogenesis of diverse lineages of dendritic cells is if they are cells originally autonomous or hold common cellular background and differentiate according to the functional environmental inputs²⁴⁸. Studies with rats^{165,41}, with mice²⁹⁷ and humans⁵¹ support the existence of diverse lineages of dendritic cells.

Dendritic cells may come from myeloid cells⁹⁸, plasmocytoid cells^{135,13}, monocytes³³⁹, macrophages^{269,187,34} or germinative blood cells^{269,302}. Some specific dendritic cells lineages hold better functional plasticity than others^{184,82,289,348} and such plasticity is exemplified by the differentiation in interdigital cells³⁴¹. Furthermore, such plasticity facilitates its collaboration in the orquestration of the immunological response¹⁶¹, presenting antigens to the T cells in a Th1 response type or inducing the host tolerance to the antigen in a Th2 response type^{194,289}.

Myeloid dendritic cells phagocytose quickly and efficiently fungus in the yeast and hyphae forms⁹⁸. Functionally, myeloid dendritic cells tend to polarize to Th1 response and are called e dendritic cells 1 (DC1). Plasmocytoid dendritic cells tend to polarize to Th2 response tolerance response, and are called dendritic cells 2 - DC2²⁶⁵. Other authors show that dendritic cells 1 may also provide Th2 responses^{247,157}, depending upon the type of the endotoxin or lypopolysaccharide as antigen and of the cytokines involved, being they type Th1 or Th2^{265,98,247,218,192}.

The acquired immunological response is triggered by the recognition of pathogens and activation of cascade events for specific inflammatory start, evolving in special toll-like receptors - TLR¹⁸⁰, considered the link between the innate and acquired immunological systems¹⁷⁷. TLR receptors are able to induce the maturation of dendritic cells and address Th1 cells responses^{318,322,167,219}; and, among such cells, the Th17 cells⁶.

LYMPHOCYTES

MHC class II molecules of the dendritic cells, in the presence of IL18 and IL12 cytokines, induce T CD4⁺ cells to Th1 acquired immunological response^{22,291}. In the absence of IL12 cytokine, the antigen presentation might induce the Th2 tolerance response²¹⁸.

The dendritic cells are helped by T CD4⁺ helper in order to present antigens to the cytotoxic CD8⁺ lymphocytes. Such help is mediated by CD40 and CD40L molecules, in the surface of T CD4⁺ helper lymphocytes. The CD40 and CD40L molecules may also be linked to other antigen-presenting cells, as macrophages and B lymphocytes²⁷⁹.

It is important to highlight that dendritic cells do not need to interact with T lymphocytes to mature⁷⁹. However, naive Th1 cells, when stimulated by DC1, present good proliferative potential and good cytotoxic power, performing important role in the acquired immunological response. Such cells produce good amount of interferon-gamma, IL2^{17,291} and IL12^{98,291}. In opposite, the naive Th2 cells, when stimulated by DC2 cells, present poor proliferative potential and poor cytotoxic power, i.e., a poor acquired immunological response. They produce good amount of IL10, TGF-beta and lower amount of interferon-gamma, do not producing IL4 or IL5. They are regulatory cells which play the immunological tolerance, expressing the role of the DC2 cells^{121,291}.

DENDRITIC CELLS AND HIV VIRUS

The HIV virus infects and replicates in dendritic cells⁴²; however, these cells maintain their capability to present antigens to T CD4⁺ cells, although such capability is depressed³⁸. Furthermore, the dendritic cells function as a vector of HIV virus infection proliferation^{119,118,245,244,61,81,319} even though they are more important as antigen-presenting cells than vectors of infection proliferation¹⁴⁴.

The infection of dendritic cells in oral mucosa of HIV+ patients might contribute to its weakness or death,⁶¹ reducing its number²³⁴. Such process also occurs in the spleen²⁰³ and in the blood^{191,127,92,230,25}.

HIV virus may also subvert the immunological system to escape its surveillance, targeting specifically C-lectin DC-SIGN receptors (DC-specific intercellular adhesion molecule-grabbing nonintegrin) of the dendritic cells³²², though interference in their intracellular signalling or their maturation inhibition and cytokines production decrease, necessary to trigger the acquired immunological response.

The dendritic cells infected by HIV virus present defect in the MHC class II molecules (as macrophages infected by HIV virus as well), that may change its ability to present antigen to CD4⁺ cells²⁴².

The oral mucosa, as the skin, does not have B lymphocytes, but only T lymphocytes, grouped in small niches randomly distributed in both sides of the basal membrane and rarely in a superficial position³²³. The oral epithelium presents approximately 37 times more T lymphocytes than the skin epithelium³²³ and the rate of lymphocytes CD4⁺/lymphocytes CD8⁺ in the oral mucosa is 1:2; in the skin is 1:4³²³, indicating that in the oral mucosa there is significantly more differentiation of CD4⁺ cells than in the skin.

The vast majority of these lymphocytes express the memory phenotype CD45RO⁺⁶³. The lymphocytes of the oral epithelium are not activated (CD25⁻), differently than the CD25⁺ lymphocytes of the adjacent connective tissue⁶³. The conversion from naive CD45A⁺ lymphocytes to memory CD45RO⁺ lymphocytes requires antigenic stimulation, suggesting that intra-epithelium apoptotic CD25⁻/CD45⁺ lymphocytes degenerate if the antigen-presenting process does not occur⁶³. CD4⁺ cells when activated differentiate in some lineages of T helper cells⁹³.

The role of CD4⁺ cells in the oral mucosa against *Candida albicans* is fundamental, although the importance of their products IL2 and interferon-gamma has not been confirmed^{147,106,105}. Other cytokines involved in such primary immunological response are IL-6 and TNF (tumor necrosis factor)-alpha¹⁰⁷. It is also possible that to occur direct antimicrobial action of T lymphocytes against *Candida* and other microorganisms¹⁷⁸.

Regulatory T cells (Treg) operate a fundamental role in the homeostasis of the immunological system^{202,30}. Basically, they control the balance between the activation and the suppression of the immunological responses, although, with such control, they limit the antipathogenic action of the host^{210,32,342,33}. The function of the Tregs is controlled by cytokines, antigen-presenting cells or directly, through TLRs (toll-like receptors) by pathogens³⁰⁴ or dendritic cells³¹ and its migration from the inflammatory site to the lymphoid site³⁴⁷. Immunoregulatory cytokines as IL10 and TGFβ, produced by innate immunological cells in response to the molecules derived from the pathogens, can be also produced by Tregs³⁰. The reduced number of Tregs in HIV+ patients suggests that such cells are lost with the HIV infection as the T conventional cells as well. However, Tregs can be preserved in lymphoid sites, and do not be infected by HIV virus, providing a partial regulatory immunological control in such different scenario^{8,224}.

T LYMPHOCYTES AND HIV VIRUS

The Th1 type acquired immunological response provided by CD4⁺ is considered the *premium* defence of the HIV+ patient against oral and vulvovaginal *candidiasis*, although the

immunological *armamentarium* against such fungal infection are complex and not totally clarified¹⁷⁴. The number of CD4⁺ lymphocytes is certainly reduced in the oral mucosa of HIV+ patients who present candidiasis^{268,112}, what is also confirmed in their periodontal tissues^{293,227}.

As alternative defence system, the epithelium of the oral mucosa induces response of the T CD8⁺ cytotoxic lymphocytes¹⁴⁷, independently of the situation of the CD4⁺ lymphocytes⁸⁵. Such CD8⁺ cells, important actors in the resistance of the oral mucosa to infections⁵⁸, are attracted to the oral epithelium¹⁸² by cytokines IL1, IL6, IL8, TNF-alpha and TGF-beta, produced by oral keratinocytes of the oral epithelium^{323,188}.

The CD8⁺ cells when activated by IL12 cytokine may inhibit the *Candida albicans* hyphae³⁵; however, it is not commonly near to the fungal hyphae because the hyphae are usually superficially located in the oral epithelium^{260,104}. The apoptosis of the CD8⁺ cells in HIV+ patients is, in general, mediated by macrophages, although CD8⁺ cells might be recruited by the oral mucosa in response to candidiasis, especially when the CD4⁺ cell number is low^{268,217}.

T lymphocytes specific for *Candida albicans*, developed by the antigenic stimulation and IL12 are eliminated in HIV+ patients, independently of its affinity degree¹⁶⁹. Furthermore, in HIV+ patients specific T lymphocytes for *Candida albicans* produce low amount of interferon-gamma, and possibly induce, by negative feedback, a Th2 tolerance response³³¹. Then, the HIV viral infection is associated to T regulatory cells (Treg) and decrease of the primary immunological response. It occurs decrease of the naive cells (CD45RO⁻) and of the memory cells (CD45RO⁺), both direct mediators of the acquired immunological response³¹¹.

In response to oral candidiasis in HIV+ patients, Th17 and IL17 cytokine are essential, offering innate and acquired immunological response throughout neutrophils and anti-microbial factors⁶⁵. T helper responses may occur throughout 03 cellular types: Th1, Th2 or Th17^{36,168}. Th17 cells come from CD4⁺ cells, and different of Th1, Th2 and Treg cells³⁰¹ e produce the IL17 cytokine. They have become the focus of the applied Immunology, because they present special functions⁹³. The role of the Th17 cells has been extensively studied *in vitro*; however, few details are known about its properties and its role in human immunological response³⁷.

In humans, Th17 cells hold distinct migratory qualities and antigenic specificity⁴. In the specific case of candidiasis, the action of Th17 cells and the cytokine IL17 have been presented of crucial importance¹⁹⁰. In the other hand, the fungal pathogenic process also holds an important role in the cellular polarization. Fungal hyphae promote the differentiation of Th17 cells and the cells Th23/cytokine IL23; however, fungal yeasts promote the differentiation of Th1 cells and the IL12 cytokine⁴. The role

of TGFβ in modulating the activation of Th17 cells is critical. Cytokines IL23, IL1 and IL6 are also involved in the antifungal defence; although their participation is not completely clear³²⁹.

In such defence, the pathogens are recognized by PRRs (pattern recognition receptors), which trigger the beginning of the immunological response to the infection²⁶⁷. The most studied way for fungus is the receptor *Dectin-1*, thru *Syk kinase* (*spleen tyrosine kinase*), *CARD9* e Raf-1¹³⁴, being critical in the induction of the Th17 cells²⁶⁷. The receptor *Dectin-1* is *C-type lectin* and is present in the NK (*natural killers*) cells, promoters of the innate response^{45,262}. In the same way, *Dectin-2*, throughout *Syk kinase* e *CARD9*, contributes for the activation of the dendritic cells and the regulation of the acquired antifungal immunological^{173,267}.

CONCLUSIONS

Oral lesions, in special the oropharyngeal opportunistic fungal infection by *Candida albicans*, have been part of the clinical evaluation of HIV+ patients and have stimulated extensive research. In such circumstances, the incidence of oral candidiasis must consider many different factors, about the fungus and about the host patient.

Currently, the main discussion in the specialized literature involves the modulation of the immunological defence in immunodepressed HIV+ patients under antiretroviral coverage. However, many aspects of the possible vulnerability of the oral mucosa and the circumstances of its breakage and the fungal colonization and invasion are not clear enough. The interaction between the host and the commensal fungus *Candida albicans* in HIV+ patients must be further explored.

REFERENCES

- [No authors listed]. Epidemiologic aspects of the current outbreak of Kaposi's sarcoma and opportunistic infections. *N Engl J Med* 306:248-252, 1982.
- [No authors listed]. Classification and diagnostic criteria for oral lesions in HIV infection. EC-Clearinghouse on Oral Problems Related to HIV Infection and WHO Collaborating Centre on Oral Manifestations of the Immunodeficiency Virus. *J Oral Pathol Med* 22:289-291, 1993.
- Abgrall S, Charreau I, Joly V, Bloch J, Reynes J, Delta Coordinating Committee. Risk factors for esophageal candidiasis in a large cohort of HIV-infected patients treated with nucleoside analogues. *Eur J Clin Microbiol Infect Dis* 20:346-349, 2001. DOI: <http://dx.doi.org/10.1007/s100960100497>
- Acosta-Rodriguez EV, Rivino L, Geginat J et al. Surface phenotype and antigenic specificity of human interleukin 17-producing T helper memory cells. *Nat Immunol* 8:639-646, 2007. PMID: 17486092 DOI: <http://dx.doi.org/10.1038/ni1467>
- Ahlfors EE, Larsson PA, Bergstresser PR. Langerhans cell surface densities in rat oral mucosa and human buccal mucosa. *J Oral Pathol* 14:390-397, 1985. PMID: 3159862 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1985.tb00510.x>

6. Allan S. Dendritic cells: tailoring T-helper-cell responses. *Nat Rev Immunol* 9:76, 2009.
7. Ammann AJ and Hong R Selective IgA deficiency: presentation of 30 cases and a review of the literature. *Medicine (Baltimore)* 50:223-236, 1971. PMID: 4938275
8. Andersson J, Boasso A, Nilsson J et al. Cutting edge: the prevalence of regulatory T cells in lymphoid tissue is correlated with viral load in HIV-infected patients. *J Immunol* 174:3143-3147, 2005.
9. Angel JB, Kumar A, Parato K et al. Improvement in cell-mediated immune function during potent anti-human immunodeficiency virus therapy with ritonavir plus saquinavir. *J Infect Dis* 177:898-904, 1998. PMID: 9534961 DOI: <http://dx.doi.org/10.1086/515244>
10. Anjuère F, Martín P, Ferrero I et al. Definition of dendritic cell subpopulations present in the spleen, Peyer's patches, lymph nodes, and skin of the mouse. *Blood* 93:590-598, 1999.
11. Ansel J, Perry P, Brown J et al. Cytokine modulation of keratinocyte cytokines. *J Investig Dermatol* 94S:101-107, 1990. DOI: <http://dx.doi.org/10.1111/1523-1747.ep12876053>
12. Antinori A, Antinori A, Ammassari A et al. Presumptive clinical criteria versus endoscopy in the diagnosis of *Candida* esophagitis at various HIV-1 disease stages. *Endoscopy* 27:371-376, 1995. PMID: 7588351 DOI: <http://dx.doi.org/10.1055/s-2007-1005716>
13. Ardavin C. Origin, precursors and differentiation of mouse dendritic cells. *Nat Rev Immunol* 3:582-590, 2003. PMID: 12876560
14. Arendorf TM and Walker DM. The prevalence and intra-oral distribution of *Candida albicans* in man. *Arch Oral Biol* 25:1-10, 1980. PMID: 6996654 DOI: [http://dx.doi.org/10.1016/0003-9969\(80\)90147-8](http://dx.doi.org/10.1016/0003-9969(80)90147-8)
15. Arribas JR, Hernández-Albujar S, González-García JJ et al. Impact of protease inhibitor therapy on HIV-related oropharyngeal candidiasis. *AIDS* 14:979-985, 2000. PMID: 10853979 DOI: <http://dx.doi.org/10.1097/00002030-200005260-00009>
16. Ashman RB. Enhancement of MHC class II antigen expression by exposure to *Candida albicans*. *Immunol Lett* 30:255-260, 1991. PMID: 1757112 DOI: [http://dx.doi.org/10.1016/0165-2478\(91\)90034-8](http://dx.doi.org/10.1016/0165-2478(91)90034-8)
17. Ashman RB and Papadimitriou JM. Production and function of cytokines in natural and acquired immunity to *Candida albicans* infection. *Microbiol Rev* 59:646-672, 1995. PMID: 8531890
18. Atkinson JC, Yeh C, Oppenheim FG, Bermudez D, Baum BJ, Fox PC. Elevation of salivary antimicrobial proteins following HIV-1 infection. *J Acquir Immune Defic Syndr* 3:41-48, 1990. PMID: 2293641
19. Autran B, Carcelain G, Li TS et al. Positive effects of combined antiretroviral therapy on CD4+ T cell homeostasis and function in advanced HIV disease. *Science* 277:112-116, 1997. PMID: 9204894 DOI: <http://dx.doi.org/10.1126/science.277.5322.112>
20. Autran B, Carcelain G, Li TS et al. Restoration of the immune system with anti-retroviral therapy. *Immunol Lett* 66:207-211, 1999. DOI: [http://dx.doi.org/10.1016/S0165-2478\(98\)00159-X](http://dx.doi.org/10.1016/S0165-2478(98)00159-X)
21. Bachmann MF and Kopf M. Balancing protective immunity and immunopathology. *Curr Opin Immunol* 14:413-419, 2002. PMID: 12088674 DOI: [http://dx.doi.org/10.1016/S0952-7915\(02\)00363-1](http://dx.doi.org/10.1016/S0952-7915(02)00363-1)
22. Banchereau J and Steinman RM. Dendritic cells and the control of immunity. *Nature* 392:245-252, 1998. PMID: 9521319 DOI: <http://dx.doi.org/10.1038/32588>
23. Barchiesi F, Arzeni D, Del Prete MS et al. Fluconazole susceptibility and strain variation of *Candida albicans* isolates from HIV-infected patients with oropharyngeal candidosis. *J Antimicrob Chemother* 41:541-548, 1998. PMID: 9630407 DOI: <http://dx.doi.org/10.1093/jac/41.5.541>
24. Barrett AW, Cruchley AT, Williams DM. Oral mucosal Langerhans' cells. *Crit Rev Oral Biol Med* 7:36-58, 1996. PMID: 8727106 DOI: <http://dx.doi.org/10.1177/10454411960070010301>
25. Barron MA, Blyveis N, Palmer BE, MaWhinney S, Wilson CC. Influence of plasma viremia on defects in number and immunophenotype of blood dendritic cell subsets in human immunodeficiency virus 1-infected individuals. *J Infect Dis* 187:26-37, 2003. PMID: 12508143 DOI: <http://dx.doi.org/10.1086/345957>
26. Basson NJ and vanWyk CW. The establishment of a community of oral bacteria that controls the growth of *Candida albicans* in a chemostat. *Oral Microbiol Immunol* 11:199-202, 1996. DOI: <http://dx.doi.org/10.1111/j.1399-302X.1996.tb00358.x>
27. Battegay M, Nüesch R, Hirschel B, Kaufmann GR. Immunological recovery and antiretroviral therapy in HIV-1 infection. *Lancet Infect Dis* 6:280-287, 2006. PMID: 16631548 DOI: [http://dx.doi.org/10.1016/S1473-3099\(06\)70463-7](http://dx.doi.org/10.1016/S1473-3099(06)70463-7)
28. Bektic J, Lell CP, Fuchs A et al. HIV protease inhibitors attenuate adherence of *Candida albicans* to epithelial cells in vitro. *FEMS Immunol Med Microbiol* 31:65-71, 2001. PMID: 11476984 DOI: [http://dx.doi.org/10.1016/S0928-8244\(01\)00242-5](http://dx.doi.org/10.1016/S0928-8244(01)00242-5)
29. Belazi M, Fleva A, Drakoulakos D, Panayiotidou D. Salivary IgA and serum IgA and IgG antibodies to *Candida albicans* in HIV-infected subjects. *Int J STD AIDS* 13:373-377, 2002. PMID: 12015010 DOI: <http://dx.doi.org/10.1258/095646202760029787>
30. Belkaid Y. Regulatory T cells and infection: a dangerous necessity. *Nat Rev Immunol* 7:875-888, 2007. PMID: 17948021
31. Belkaid Y and Oldenhove G. Tuning microenvironments: induction of regulatory T cells by dendritic cells. *Immunity* 29:362-371, 2008. PMID: 18799144
32. Belkaid Y and Rouse BT. Natural regulatory T cells in infectious disease. *Nat Immunol* 6:353-360, 2005. PMID: 15785761 DOI: <http://dx.doi.org/10.1038/ni1181>
33. Belkaid Y and Tarbell K. Regulatory T cells in the control of host-microorganism interactions. *Ann Rev Immunol* 27:551-589, 2009. DOI: <http://dx.doi.org/10.1146/annurev.immunol.021908.132723>
34. Bender A, Sapp M, Schuler G, Steinman RM, Bhardwaj N. Improved methods for the generation of dendritic cells from nonproliferating progenitors in human blood. *J Immunol Methods* 196:121-135, 1996. DOI: [http://dx.doi.org/10.1016/0022-1759\(96\)00079-8](http://dx.doi.org/10.1016/0022-1759(96)00079-8)
35. Beno DW, Stöver AG, Mathews HL. Growth inhibition of *Candida albicans* hyphae by CD8+ lymphocytes. *J Immunol* 154:5273-5281, 1995. PMID: 7730631
36. Bettelli E, Korn T, Kuchroo VK. Th17: the third member of the effector T cell trilogy. *Curr Opin Immunol* 19:652-657, 2007. PMID: 17766098
37. Bird L. T cells: human Th17 cells take centre stage. *Nat Rev Immunol* 7:413, 2007.
38. Blauvelt A, Clerici M, Lucey DR et al. Functional studies of epidermal Langerhans cells and blood monocytes in HIV-infected persons. *J Immunol* 154:3506-3515, 1995.
39. Borg M and Rüchel R. Expression of extracellular acid proteinase by proteolytic *Candida* spp. during experimental infection of oral mucosa. *Infect Immun* 56:626-631, 1988. PMID: 3277916
40. Bos IR and Burkhardt A. Interepithelial cells of the oral mucosa. Light and electron microscopic observations in germfree, specific pathogen-free and conventionalized mice. *J Oral Pathol* 9:65-81, 1980. PMID: 6768863 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1980.tb01389.x>

41. Bowers WE and Berkowitz MR. Differentiation of dendritic cells in cultures of rat bone marrow cells. *J Exp Med* 163:872-883, 1986. PMID: 3512761 DOI: <http://dx.doi.org/10.1084/jem.163.4.872>
42. Braathen LR, Ramirez G, Kunze RO, Gelderblom H. Langerhans cells as primary target cells for HIV infection. *Lancet* 2:1094, 1987. PMID: 2890007 DOI: [http://dx.doi.org/10.1016/S0140-6736\(87\)91526-1](http://dx.doi.org/10.1016/S0140-6736(87)91526-1)
43. Brambilla AM, Castagna A, Nocita B et al. Relation between CD4 cell counts and HIV RNA levels at onset of opportunistic infections. *J Acquir Immune Syndr* 27:44-48, 2001. PMID: 11404519
44. Brodt HR, Kamps BS, Gute P, Knupp B, Staszewski S, Helm EB. Changing incidence of AIDS-defining illnesses in the era of antiretroviral combination therapy. *AIDS* 11:1731-1738, 1997. PMID: 9386808 DOI: <http://dx.doi.org/10.1097/00002030-199714000-00010>
45. Brown GD. Dectin-1: a signalling non-TLR pattern-recognition receptor. *Nat Rev Immunol* 6:33-43, 2006.
46. Campo J, Del Romero J, Castilla J, Garcia S, Rodriguez C, Bascones A. Oral candidiasis as a clinical marker related to viral load, CD4 lymphocyte count and CD4 lymphocyte percentage in HIV-infected patients. *J Oral Pathol Med* 31:5-10, 2002. PMID: 11896816 DOI: <http://dx.doi.org/10.1034/j.1600-0714.2002.310102.x>
47. Cantorna MT and Balish E. Role of CD4+ lymphocytes in resistance to mucosal candidiasis. *Infect Immun* 59:2447-2455, 1991. PMID: 1675629
48. Cassone A, de Bernardis F, Torosantucci A, Tacconelli E, Tumbarello M, Cauda R. In vitro and in vivo anticandidal activity of human immunodeficiency virus protease inhibitors. *J Infect Dis* 180:448-453, 1999. DOI: <http://dx.doi.org/10.1086/314871>
49. Cassone A, Tacconelli E, de Bernardis F et al. Antiretroviral therapy with protease inhibitors has an early, immune reconstitution-independent beneficial effect on *Candida* virulence and oral candidiasis in human immunodeficiency virus-infected subjects. *J Infect Dis* 185:188-195, 2002. PMID: 11807692 DOI: <http://dx.doi.org/10.1086/338445>
50. Cauda R, Tacconelli E, Tumbarello M et al. Role of protease inhibitors in preventing recurrent oral candidosis in patients with HIV infection: a prospective case-control study. *J Acquir Immune Defic Syndr* 21:20-25, 1999. PMID: 10235510
51. Caux C, Dezutter-Dambuyant C, Schmitt D, Banchereau J. GM-CSF and TNF-alpha cooperate in the generation of dendritic Langerhans cells. *Nature* 360:258-261, 1992. PMID: 1279441 DOI: <http://dx.doi.org/10.1038/360258a0>
52. Caux C, Massacrier C, Vanbervliet B et al. CD34+ hematopoietic progenitors from human cord blood differentiate along two independent dendritic cell pathways in response to granulocyte-macrophage colony-stimulating factor plus tumor necrosis factor alpha: II. Functional analysis. *Blood* 90:1458-1470, 1997. PMID: 9269763
53. Caux C, Vanbervliet B, Massacrier C et al. CD34+ hematopoietic progenitors from human cord blood differentiate along two independent dendritic cell pathways in response to GM-CSF+TNF alpha. *J Exp Med* 184:695-706, 1996. PMID: 8760823 DOI: <http://dx.doi.org/10.1084/jem.184.2.695>
54. Cella M, Sallusto F, Lanzavecchia A. Origin, maturation and antigen presenting function of dendritic cells. *Curr Opin Immunol* 9:10-16, 1997. PMID: 9039784 DOI: [http://dx.doi.org/10.1016/S0952-7915\(97\)80153-7](http://dx.doi.org/10.1016/S0952-7915(97)80153-7)
55. Cilla G, Perez TE, Furundarena JR, Cuadrado E, Iribarren JA, Neira F. Esophageal candidiasis and immunodeficiency associated with acute HIV infection. *AIDS* 2:399-400, 1988.
56. Challacombe SJ. Immunologic aspects of oral candidiasis. *Oral Surg Oral Med Oral Pathol* 78:202-210, 1994. PMID: 7936590 DOI: [http://dx.doi.org/10.1016/0030-4220\(94\)90148-1](http://dx.doi.org/10.1016/0030-4220(94)90148-1)
57. Challacombe SJ and Naglik JR. The effects of HIV infection on oral mucosal immunity. *Adv Dent Res* 19:29-35, 2006. PMID: 16672546 DOI: <http://dx.doi.org/10.1177/154407370601900107>
58. Challacombe SJ and Sweet SP. Oral mucosal immunity and HIV infection: current status. *Oral Dis* 8S:55-62, 2002.
59. Chandra J, Kuhn DM, Mukherjee PK, Hoyer LL, McCormick T, Ghannoum MA. Biofilm formation by the fungal pathogen *Candida albicans*: development, architecture, and drug resistance. *J Bacteriol* 183:5385-5394, 2001. PMID: 11514524
60. Chiou CC, Groll AH, Gonzalez CE et al. Esophageal candidiasis in pediatric acquired immunodeficiency syndrome: clinical manifestations and risk factors. *Pediatr Infect Dis J* 19:729-734, 2000. PMID: 10959741
61. Chou LL, Epstein J, Cassol AS, West DM, He W, Firth JD. Oral mucosal Langerhans' cells as target, effector and vector in HIV infection. *J Oral Pathol Med* 29:394-402, 2000. DOI: <http://dx.doi.org/10.1034/j.1600-0714.2000.290805.x>
62. Clerici M and Shearer GMA. TH1→TH2 switch is a critical step in the etiology of HIV infection. *Immunol Today* 14:107-111, 1993. PMID: 8096699 DOI: [http://dx.doi.org/10.1016/0167-5699\(93\)90208-3](http://dx.doi.org/10.1016/0167-5699(93)90208-3)
63. Colasante A, Rosini S, Piattelli A, Artese L, Aiello FB, Musiani P. Distribution and phenotype of immune cells in normal human gingiva: active immune response versus unresponsiveness. *J Oral Pathol Med* 21:12-16, 1992. PMID: 1534371
64. Coleman D, Sullivan D, Harrington B et al. Molecular and phenotypic analysis of *Candida dubliniensis*: a recently identified species linked with oral candidosis in HIV-infection and AIDS patients. *Oral Dis* 3S:96-101, 1997.
65. Conti HR, Shen F, Namrata N et al. Th17 cells and IL-17 receptor signaling are essential for mucosal host defense against oral candidiasis. *J Exp Med* 206:299-311, 2009. DOI: <http://dx.doi.org/10.1084/jem.20081463>
66. Coogan MM and Challacombe SJ. Serum and salivary antibodies to a mycobacterial 65-kDa stress protein are elevated in HIV-positive patients and modified by oral candidiasis. *Oral Microbiol Immunol* 15:284-289, 2000. PMID: 11154418 DOI: <http://dx.doi.org/10.1034/j.1399-302x.2000.150503.x>
67. Coogan MM, Greenspan JS, Challacombe SJ. Oral lesions in infection with human immunodeficiency virus. *Bull World Health Organ* 83:700-706, 2005.
68. Coogan MM, Sweet SP, Challacombe SJ. Immunoglobulin A (IgA), IgA1, and IgA2 antibodies to *Candida albicans* in whole and parotid saliva in human immunodeficiency virus infection and AIDS. *Infect Immun* 62:892-896, 1994. PMID: 8112860
69. Cruchley AT, Williams DM, Farthing PM, Lesch CA, Squier CA. Regional variation in Langerhans cell distribution and density in normal human oral mucosa determined using monoclonal antibodies against CD1, HLADR, HLADQ and HLADP. *J Oral Pathol Med* 18:510-516, 1989. PMID: 2481737 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1989.tb01353.x>

70. Cutler JE. Putative virulence factors of *Candida albicans*. *Annu Rev Microbiol* 45:187-218, 1991. PMID: 1741614 DOI: <http://dx.doi.org/10.1146/annurev.micro.45.1.187>
71. Cutler CW, Jotwani R, Pulerdran B. Dendritic cells: immune saviors or Achilles' heel? *Infect Immun* 69:4703-4708, 2001. DOI: <http://dx.doi.org/10.1128/IAI.69.8.4703-4708.2001>
72. Dahl KM, Keath EJ, Fraser VJ, Powderly WG. Molecular epidemiology of mucosal candidiasis in HIV-positive women. *AIDS Res Hum Retroviruses* 13:485-491, 1997. PMID: 9100990 DOI: <http://dx.doi.org/10.1089/aid.1997.13.485>
73. Daniels TE. Human mucosal Langerhans cells: postmortem identification of regional variations in oral mucosa. *J Invest Dermatol* 82:21-24, 1984. PMID: 6228611
74. Darbyshire J. Therapeutic interventions in HIV infection - a critical view. *Trop Med Int Health* 5A:26-31, 2000. DOI: <http://dx.doi.org/10.1046/j.1365-3156.2000.00597.x>
75. da Silva CA, Dourado I, Dahia SR, Harzheim E, Rutherford GW. Oral manifestations of HIV infection in patients receiving highly active antiretroviral therapy (HAART) in Bahia, Brazil. *J Public Health Dent* 68:178-181, 2008. DOI: <http://dx.doi.org/10.1111/j.1752-7325.2007.00071.x>
76. de Bernardis F, Chiani P, Ciccozzi M et al. Elevated aspartic proteinase secretion and experimental pathogenicity of *Candida albicans* isolates from oral cavities of subjects infected with human immunodeficiency virus. *Infect Immun* 64:466-471, 1996. PMID: 8550193
77. de Bernardis F, Mühlischlegel FA, Cassone A, Fonzi WA. The pH of the host niche controls gene expression in and virulence of *Candida albicans*. *Infect Immun* 66:3317-3325, 1998.
78. de Bernardis F, Sullivan PA, Cassone A. Aspartyl proteinases of *Candida albicans* and their role in pathogenicity. *Med Mycol* 39:303-313, 2001. PMID: 11556759
79. de Creus A, van Beneden K, Taghon T et al. Langerhans cells that have matured in vivo in the absence of T cells are fully capable of inducing a helper CD4 as well as a cytotoxic CD8 response. *J Immunol* 165:645-653, 2000.
80. de Repentigny L. Serodiagnosis of candidiasis, aspergillosis, and cryptococcosis. *Clin Infect Dis* 14S:11-22, 1992. DOI: http://dx.doi.org/10.1093/clinids/14.Supplement_1.S11
81. de Repentigny L, Lewandowski D, Jolicouer P. Immunopathogenesis of oropharyngeal candidiasis in human immunodeficiency virus infection. *Clin Microbiol Rev* 17:729-759, 2004. DOI: <http://dx.doi.org/10.1128/CMR.17.4.729-759.2004>
82. del Hoyo GM, Martín P, Vargas HH, Ruiz S, Arias CF, Ardavin C. Characterization of a common precursor population for dendritic cells. *Nature* 415:1043-1047, 2002. DOI: <http://dx.doi.org/10.1038/4151043a>
83. Deeks SG, Smith M, Holodniy M, Kahn JO. HIV-1 protease inhibitors. A review for clinicians. *JAMA* 277:145-153, 1997. PMID: 8990341 DOI: <http://dx.doi.org/10.1001/jama.277.2.145>
84. de Repentigny L, Lewandowski D, Jolicouer P. Immunopathogenesis of oropharyngeal candidiasis in human immunodeficiency virus infection. *Clin Microbiol Rev* 17:729-759, 2004. PMID: 15489345 DOI: <http://dx.doi.org/10.1128/CMR.17.4.729-759.2004>
85. Desvignes C, Estèves F, Etchart N, Bella C, Czerkinsky C, Kaiserlian D. The murine buccal mucosa is an inductive site for priming class I-restricted CD8+ effector T cells in vivo. *Clin Exp Immunol* 113:386-393, 1998. DOI: <http://dx.doi.org/10.1046/j.1365-2249.1998.00671.x>
86. Detels R, Tarwater P, Phair JP et al. Effectiveness of potent antiretroviral therapies on the incidence of opportunistic infections before and after AIDS diagnosis. *AIDS* 15:347-355, 2001. PMID: 11273215 DOI: <http://dx.doi.org/10.1097/00002030-200102160-00008>
87. Díaz-Guerra TM, Martínez-Suárez JV, Laguna F, Rodríguez-Tudela JL. Comparison of four molecular typing methods for evaluating genetic diversity among *Candida albicans* isolates from human immunodeficiency virus-positive patients with oral candidiasis. *J Clin Microbiol* 35:856-861, 1997. PMID: 9157142
88. Dieleman JP, Jambroes M, Gyssens IC et al. Determinants of recurrent toxicity-driven switches of highly active antiretroviral therapy. The ATHENA cohort. *AIDS* 16:737-745, 2002. PMID: 11964530 DOI: <http://dx.doi.org/10.1097/00002030-200203290-00009>
89. Diz Dios P, Ocampo A, Miralles C, Otero I, Iglesias I, Rayo N. Frequency of oropharyngeal candidiasis in HIV-infected patients on protease inhibitor therapy. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 87:437-441, 1999. PMID: 10225625 DOI: [http://dx.doi.org/10.1016/S1079-2104\(99\)70242-8](http://dx.doi.org/10.1016/S1079-2104(99)70242-8)
90. Diz Dios P, Ocampo A, Otero I, Iglesias I, Martínez C. Changes in oropharyngeal colonization and infection by *Candida albicans* in human immunodeficiency virus-infected patients. *J Infect Dis* 183:355-356, 2001. PMID: 11120936
91. Dodd CL, Greenspan D, Katz MH, Westenhouse JL, Feigal DW, Greenspan JS. Oral candidiasis in HIV infection: pseudomembranous and erythematous candidiasis show similar rates of progression to AIDS. *AIDS* 5:1339-1343, 1991. PMID: 1768382
92. Donaghy H, Pozniak A, Gazzard B et al. Loss of blood CD11c+ myeloid and CD11c- plasmacytoid dendritic cells in patients with HIV-1 infection correlates with HIV-1 RNA virus load. *Blood* 98:2574-2576, 2001. PMID: 11588058 DOI: <http://dx.doi.org/10.1182/blood.V98.8.2574>
93. Dong C. Th17 cells in development: an updated view of their molecular identity and genetic programming. *Nat Rev Immunol* 8:337-348, 2008. PMID: 18408735
94. Dongari-Bagtzoglou A and Fidel PL Jr. The host cytokine responses and protective immunity in oropharyngeal candidiasis. *J Dent Res* 84:966-977, 2005. PMID: 16246925 DOI: <http://dx.doi.org/10.1177/154405910508401101>
95. Dongari-Bagtzoglou A and Kashleva H. Granulocyte-macrophage colony-stimulating factor responses of oral epithelial cells to *Candida albicans*. *Oral Microbiol Immunol* 18:165-170, 2003. PMID: 12753468 DOI: <http://dx.doi.org/10.1034/j.1399-302X.2003.00061.x>
96. Dongari-Bagtzoglou A, Kashleva H, Villar CC. Bioactive interleukin-1 alpha is cytolytically released from *Candida albicans*-infected oral epithelial cells. *Med Mycol* 42:531-541, 2004. PMID: 15682642
97. Dorhoi A and Kaufmann SH. Fine-tuning of T cell responses during infection. *Curr Opin Immunol* 21:367-377, 2009. PMID: 19646852 DOI: <http://dx.doi.org/10.1016/j.coi.2009.07.004>
98. d'Ostiani CF, Del Sero G, Bacci A et al. Dendritic cells discriminate between yeasts and hyphae of the fungus *Candida albicans*. Implications for initiation of T helper cell immunity in vitro and in vivo. *J Exp Med* 191:1661-1674, 2000.
99. Drobacheff C, Millon L, Monod M et al. Increased serum and salivary immunoglobulins against *Candida albicans* in HIV-infected patients with oral candidiasis. *Clin Chem Lab Med* 39:519-526, 2001. PMID: 11506465 DOI: <http://dx.doi.org/10.1515/CCLM.2001.087>

100. Edgerton M, Koshlukova SE, Lo TE, Chrzan BG, Straubinger RM, Raj PA. Candidacidal activity of salivary histatins. Identification of a histatin 5-binding protein on *Candida albicans*. *J Biol Chem* 273:20438-20447, 1998. DOI: <http://dx.doi.org/10.1074/jbc.273.32.20438>
101. Edgerton M, Scannapieco FA, Reddy MS, Levine MJ. Human submandibular-sublingual saliva promotes adhesion of *Candida albicans* to polymethylmethacrylate. *Infect Immun* 61:2644-2652, 1993. PMID: 8500903
102. Elahi S, Pang G, Clancy R, Ashman RB. Cellular and cytokine correlates of mucosal protection in murine model of oral candidiasis. *Infect Immun* 68:5771-5777, 2000. PMID: 10992484 DOI: <http://dx.doi.org/10.1128/IAI.68.10.5771-5777.2000>
103. Epstein JB, Kimura LH, Menard TW, Truelove EL, Pearsall NN. Effects of specific antibodies on the interaction between the fungus *Candida albicans* and human oral mucosa. *Arch Oral Biol* 27:469-474, 1982. PMID: 6956259 DOI: [http://dx.doi.org/10.1016/0003-9969\(82\)90086-3](http://dx.doi.org/10.1016/0003-9969(82)90086-3)
104. Eversole LR, Reichart PA, Ficarra G, Schmidt-Westhausen A, Romagnoli P, Pimpinelli N. Oral keratinocyte immune responses in HIV-associated candidiasis. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 84:372-380, 1997. PMID: 9347501 DOI: [http://dx.doi.org/10.1016/S1079-2104\(97\)90035-4](http://dx.doi.org/10.1016/S1079-2104(97)90035-4)
105. Farah CS, Elahi S, Drysdale K et al. Primary role for CD4+ T lymphocytes in recovery from oropharyngeal candidiasis. *Infect Immun* 70:724-731, 2002. PMID: 11796605 DOI: <http://dx.doi.org/10.1128/IAI.70.2.724-731.2002>
106. Farah CS, Elahi S, Pang G et al. T cells augment monocyte and neutrophil function in host resistance against oropharyngeal candidiasis. *Infect Immun* 69:6110-6118, 2001. PMID: 11553549 DOI: <http://dx.doi.org/10.1128/IAI.69.10.6110-6118.2001>
107. Farah CS, Gotjamanos T, Seymour GJ, Ashman RB. Cytokines in the oral mucosa of mice infected with *Candida albicans*. *Oral Microbiol Immunol* 17:375-378, 2002. PMID: 12485329 DOI: <http://dx.doi.org/10.1034/j.1399-302X.2002.170607.x>
108. Feigal DW, Katz MH, Greenspan D et al. The prevalence of oral lesions in HIV-infected homosexual and bisexual men: three San Francisco epidemiological cohorts. *AIDS* 5:519-525, 1991. PMID: 1863403
109. Ferreira S, Noce C, Junior AS et al. Prevalence of oral manifestations of HIV infection in Rio de Janeiro, Brazil from 1988 to 2004. *AIDS Patient Care STDS* 21:724-731, 2007. PMID: 17949271 DOI: <http://dx.doi.org/10.1089/apc.2006.0211>
110. Fichtenbaum CJ, Koletar S, Yiannoutsos C et al. Refractory mucosal candidiasis in advanced human immunodeficiency virus infection. *Clin Infect Dis* 30:749-756, 2000. PMID: 10816143 DOI: <http://dx.doi.org/10.1086/313765>
111. Fidel PL Jr. Distinct protective host defenses against oral and vaginal candidiasis. *Med Mycol* 40:359-375, 2002. PMID: 12230215
112. Fidel PL Jr. Immunity to *Candida*. *Oral Dis* 8S:69-75, 2002. DOI: <http://dx.doi.org/10.1034/j.1601-0825.2002.00015.x>
113. Fidel PL Jr. and Sobel JD. Immunopathogenesis of recurrent vulvovaginal candidiasis. *Clin Microbiol Rev* 9:335-348, 1996. PMID: 8809464
114. Fidel PL Jr., Vazquez JA, Sobel JD. *Candida glabrata*: review of epidemiology, pathogenesis, and clinical disease with comparison to *C. albicans*. *Clin Microbiol Rev* 12:80-96, 1999.
115. Fidel PL Jr., Wolf NA, KuKuruga MA. T lymphocytes in the murine vaginal mucosa are phenotypically distinct from those in the periphery. *Infect Immun* 64:3793-3799, 1996. PMID: 8751931
116. Fong IW, Laurel M, Burford-Mason A. Asymptomatic oral carriage of *Candida albicans* in patients with HIV infection. *Clin Investig Med* 20:85-93, 1997.
117. Formanek M, Knerer B, Kornfehl J. Cytokine expression of human oral keratinocytes. *ORL J Otorhinolaryngol Relat Spec* 61:103-107, 1999. PMID: 10095201 DOI: <http://dx.doi.org/10.1159/000027650>
118. Frankel SS, Tenner-Racz K, Racz P et al. Active replication of HIV-1 at the lymphoepithelial surface of the tonsil. *Am J Pathol* 151:89-96, 1997. PMID: 9212735
119. Frankel SS, Wenig BM, Burke AP et al. Replication of the HIV-1 in dendritic cell-derived syncytia at the mucosal surface of the adenoid. *Science* 272:115-117, 1996. PMID: 8600520 DOI: <http://dx.doi.org/10.1126/science.272.5258.115>
120. Gillespie GM and Mariño R. Oral manifestations of HIV infection: a Panamerican perspective. *J Oral Pathol Med* 22:2-7, 1993.
121. Gilliet M and Liu YJ. Generation of human CD8 T regulatory cells by CD40 ligand-activated plasmacytoid dendritic cells. *J Exp Med* 195:695-704, 2002. PMID: 11901196 DOI: <http://dx.doi.org/10.1084/jem.20011603>
122. Giuliani M, Lajolo C, Sartorio A et al. Oral lesions in HIV and HCV co-infected individuals in HAART era. *J Oral Pathol Med* 37:468-474, 2008. PMID: 18298476 DOI: <http://dx.doi.org/10.1111/j.1600-0714.2008.00647.x>
123. Ghannoum MA. Potential role of phospholipases in virulence and fungal pathogenesis. *Clin Microbiol Rev* 13:122-143, 2000. PMID: 10627494 DOI: <http://dx.doi.org/10.1128/CMR.13.1.122-143.2000>
124. Glick M, Muzyka BC, Lurie D, Salkin LM. Oral manifestations associated with HIV-related disease as markers for immune suppression and AIDS. *Oral Surg Oral Med Oral Pathol* 77:344-349, 1994. PMID: 8015797 DOI: [http://dx.doi.org/10.1016/0030-4220\(94\)90195-3](http://dx.doi.org/10.1016/0030-4220(94)90195-3)
125. Goupil M, Trudelle EB, Dugas V et al. Macrophage-mediated responses to *Candida albicans* in mice expressing the human immunodeficiency virus type 1 transgene. *Infect Immunol* 77:4136-4149, 2009. DOI: <http://dx.doi.org/10.1128/IAI.00453-09>
126. Gottlieb MS, Schroff R, Schanker HM et al. Pneumocystis carinii pneumonia and mucosal candidiasis in previously healthy homosexual men: evidence of a new acquired cellular immunodeficiency. *N Engl J Med* 305:1425-1431, 1981.
127. Grassi F, Hosmalin A, McLroy D, Calvez V, Debré P, Autran B. Depletion in blood CD11c-positive dendritic cells from HIV-infected patients. *AIDS* 13:759-766, 1999. PMID: 10357374 DOI: <http://dx.doi.org/10.1097/00002030-199905070-00004>
128. Greenspan JS, Barr CE, Sciubba JJ, Winkler JR. Oral manifestations of HIV infection. Definitions, diagnostic criteria, and principles of therapy. The USA Oral AIDS Collaborative Group. *Oral Surg Oral Med Oral Pathol* 73:142-144, 1992. DOI: [http://dx.doi.org/10.1016/0030-4220\(92\)90185-S](http://dx.doi.org/10.1016/0030-4220(92)90185-S)
129. Greenspan D, Canchola AJ, MacPhail LA, Cheikh B, Greenspan JS. Effect of highly active antiretroviral therapy on frequency of oral warts. *Lancet* 357:1411-1412, 2001. PMID: 11356441 DOI: [http://dx.doi.org/10.1016/S0140-6736\(00\)04578-5](http://dx.doi.org/10.1016/S0140-6736(00)04578-5)

130. Greenspan D and Greenspan JS. Oral manifestations of human immunodeficiency virus infection. *Dent Clin North Am* 37:21-32, 1993. PMID: 8416823
131. Greenspan D and Greenspan JS. HIV-related oral disease. *Lancet* 348:729-733, 1996. PMID: 8806295 DOI: [http://dx.doi.org/10.1016/S0140-6736\(96\)02308-2](http://dx.doi.org/10.1016/S0140-6736(96)02308-2)
132. Greenspan D, Greenspan JS, Conant M, Petersen V, Silverman S Jr., de Souza Y. Oral "hairy" leucoplakia in male homosexuals: evidence of association with both papillomavirus and a herpes-group virus. *Lancet* 2:831-834, 1984. PMID: 6148571
133. Greenspan D, Komaroff E, Redford M et al. Oral mucosal lesions and HIV viral load in the Women's Interagency HIV Study (WIHS). *J Acquir Immune Defic Syndr* 25:44-50, 2000. PMID: 11064503 DOI: <http://dx.doi.org/10.1097/00126334-200009010-00006>
134. Gringhuis SI, den Dunnen J, Litjens M et al. Dectin-1 directs T helper cell differentiation by controlling noncanonical NF- κ B activation through Raf-1 and Syk. *Nat Immunol* 10:203-213, 2009. PMID: 19122653 DOI: <http://dx.doi.org/10.1038/ni.1692>
135. Grouard G, Risoan MC, Filgueira L, Durand I, Banchereau J, Liu YJ. The enigmatic plasmacytoid T cells develop into dendritic cells with interleukin (IL)-3 and CD40-ligand. *J Exp Med* 185:1101-1111, 1997. PMID: 9091583
136. Gruber A, Lukasser-Vogl E, Borg-von Zepelin M, Dierich MP, Würzner R. Human immunodeficiency virus type 1 gp160 and gp41 binding to *Candida albicans* selectively enhances candidal virulence in vitro. *J Infect Dis* 177:1057-1063, 1998. PMID: 9534982 DOI: <http://dx.doi.org/10.1086/515231>
137. Hacker MA, Kaida A, Hogg RS, Bastos FI. The first ten years: achievements and challenges of the Brazilian program of universal access to HIV/AIDS comprehensive management and care, 1996-2006. *Cad Saude Publica* 23S:345-359, 2007.
138. Hasséus B, Jontell M, Bergenholtz G, Eklund C, Dahlgren UI. Langerhans cells from oral epithelium are more effective in stimulating allogeneic T-cells in vitro than Langerhans cells from skin epithelium. *J Dent Res* 78:751-758, 1999. PMID: 10096450
139. Helmerhorst EJ, Flora B, Troxler RF, Oppenheim FG. Dialysis unmasks the fungicidal properties of glandular salivary secretions. *Infect Immun* 72:2703-2709, 2004. PMID: 15102779 DOI: <http://dx.doi.org/10.1128/IAI.72.5.2703-2709.2004>
140. Hibino K, Samaranyake LP, Hägg U, Wong RWK, Lee W. The role of salivary factors in persistent oral carriage of *Candida* in humans. *Arch Oral Biol* 54:678-683, 2009. DOI: <http://dx.doi.org/10.1016/j.archoralbio.2009.04.003>
141. Hill IR and Porter P. Studies of bactericidal activity to *Escherichia coli* of porcine serum and colostral immunoglobulins and the role of lysozyme with secretory IgA. *Immunology* 26:1239-1250, 1974.
142. Hodgson TA, Greenspan D, Greenspan JS. Oral lesions of HIV disease and HAART in industrialized countries. *Adv Dent Res* 19:57-62, 2006. PMID: 16672551 DOI: <http://dx.doi.org/10.1177/154407370601900112>
143. Hoffman MP and Haidaris CG. Analysis of *Candida albicans* adhesion to salivary mucin. *Infect Immun* 61:1940-1949, 1993. PMID: 8478083
144. Hogue IB, Bajaria SH, Fallert BA, Qin S, Reinhart TA, Kirschner DE. The dual role of dendritic cells in the immune response to human immunodeficiency virus type 1 infection. *J Gen Virol* 89:2228-2239, 2008. PMID: 18753232 DOI: <http://dx.doi.org/10.1099/vir.0.83600-0>
145. Hohl TM, Rivera A, Palmer EG. Immunity to fungi. *Curr Opin Immunol* 18:465-472, 2006. DOI: <http://dx.doi.org/10.1016/j.coi.2006.05.003>
146. Holmberg K and Meyer RD. Fungal infections in patients with AIDS and AIDS-related complex. *Scand J Infect Dis* 18:179-192, 1986. PMID: 3526530 DOI: <http://dx.doi.org/10.3109/00365548609032326>
147. Howcroft TK, Strebel K, Martin MA, Singer DS. Repression of MHC class I gene promoter activity by two-exon Tat of HIV. *Science* 260:1320-1322, 1993. PMID: 8493575 DOI: <http://dx.doi.org/10.1126/science.8493575>
148. Hube B. *Candida albicans* secreted aspartyl proteinases. *Curr Top Med Mycol* 7:55-69, 1996. PMID: 9504059
149. Hube B, Monod M, Schofield DA, Brown AJ, Gow NA. Expression of seven members of the gene family encoding secretory aspartyl proteinases in *Candida albicans*. *Mol Microbiol* 14:87-99, 1994. PMID: 7830564 DOI: <http://dx.doi.org/10.1111/j.1365-2958.1994.tb01269.x>
150. Hussain LA and Lehner T. Comparative investigation of Langerhans' cells and potential receptors for HIV in oral, genitourinary and rectal epithelia. *Immunology* 85:475-484, 1995. PMID: 7558138
151. Ibrahim AS, Mirbod F, Filler SG et al. Evidence implicating phospholipase as a virulence factor of *Candida albicans*. *Infect Immun* 63:1993-1998, 1995. PMID: 7729913
152. Ives NJ, Gazzard BG, Easterbrook PJ. The changing pattern of AIDS-defining illnesses with the introduction of highly active antiretroviral therapy (HAART) in a London clinic. *J Infect* 42:134-139, 2001. DOI: <http://dx.doi.org/10.1053/jinf.2001.0810>
153. Iwazaki A. Mucosal dendritic cells. *Annu Rev Immunol* 25:381-418, 2007.
154. Janeway CA, Jr. The immune system evolved to discriminate infectious nonself from infectious self. *Immunol Today* 13:11-16, 1992. DOI: [http://dx.doi.org/10.1016/0167-5699\(92\)90198-G](http://dx.doi.org/10.1016/0167-5699(92)90198-G)
155. Jones-Carson J, Vásquez-Torres A, van der Heyde HC, Warner T, Wagner RD, Balish E. Gamma delta T cell-induced nitric oxide production enhances resistance to mucosal candidiasis. *Nat Med* 1:552-557, 1995. DOI: <http://dx.doi.org/10.1038/nm0695-552>
156. Jonuleit H, Schmitt E, Schuler G, Knop J, Enk AH. Induction of interleukin 10-producing, nonproliferating CD4+ T cells with regulatory properties by repetitive stimulation with allogeneic immature human dendritic cells. *J Exp Med* 192:1213-1222, 2000. PMID: 11067871 DOI: <http://dx.doi.org/10.1084/jem.192.9.1213>
157. Jotwani R and Cutler CW. Multiple dendritic cell (DC) subpopulations in human gingiva and association of mature dendritic cells with CD4+ T-cells in situ. *J Dent Res* 82:736-741, 2003. DOI: <http://dx.doi.org/10.1177/154405910308200915>
158. Jotwani R, Palucka AK, Al Quotub M et al. Mature dendritic cells infiltrate the T cell-rich region of oral mucosa in chronic periodontitis: in situ, in vivo, and in vitro studies. *J Immunol* 167:4693-4700, 2001.
159. Kalinski P and Moser M. Consensual immunity: success-driven development of T-helper-1 and T-helper-2 responses. *Nat Rev Immunol* 5:252-260, 2005.
160. Kaplan JE, Hanson D, Dworkin MS et al. Epidemiology of human immunodeficiency virus-associated opportunistic infections in the United States in the era of highly active antiretroviral therapy. *Clin Infect Dis* 30S:5-14, 2000. DOI: <http://dx.doi.org/10.1086/313843>

161. Kapsenberg ML. Dendritic-cell control of pathogen-driven T-cell polarization. *Nat Rev Immunol* 3:984-993, 2003. PMID: 14647480 DOI: <http://dx.doi.org/10.1038/nri1246>
162. Katz MH, Greenspan D, Westenhoe J et al. Progression to AIDS in HIV-infected homosexual and bisexual men with hairy leukoplakia and oral candidiasis. *AIDS* 6:95-100, 1992. PMID: 1543572 DOI: <http://dx.doi.org/10.1097/00002030-199201000-00013>
163. Kirby AJ, Muñoz A, Detels R, Armstrong JA, Saah A, Phair JP. Thrush and fever as measures of immunocompetence in HIV-1-infected men. *J Acquir Immune Defic Syndr* 7:1242-1249, 1994. DOI: <http://dx.doi.org/10.1097/00126334-199412000-00005>
164. Klein RS, Harris CA, Small CB, Moll B, Lesser M, Friedland GH. Oral candidiasis in high-risk patients as the initial manifestation of the acquired immunodeficiency syndrome. *N Engl J Med* 311:354-358, 1984. PMID: 6738653 DOI: <http://dx.doi.org/10.1056/NEJM198408093110602>
165. Klinkerk WE. Rat bone marrow precursors develop into dendritic accessory cells under the influence of a conditioned medium. *Immunobiology* 168:414-424, 1984.
166. Kolokotronis A, Kioses V, Antoniadis D, Mandraveli K, Doutsos I, Papanayotou P. Immunologic status in patients infected with HIV with oral candidiasis and hairy leukoplakia. *Oral Surg Oral Med Oral Pathol* 78:41-46, 1994. PMID: 8078662 DOI: [http://dx.doi.org/10.1016/0030-4220\(94\)90115-5](http://dx.doi.org/10.1016/0030-4220(94)90115-5)
167. Kopp E and Medzhitov R. Recognition of microbial infection by Toll-like receptors. *Curr Opin Immunol* 15:396-401, 2003. PMID: 12900270 DOI: [http://dx.doi.org/10.1016/S0952-7915\(03\)00080-3](http://dx.doi.org/10.1016/S0952-7915(03)00080-3)
168. Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 cells. *Annu Rev Immunol* 27:485-517, 2009. PMID: 19132915 DOI: <http://dx.doi.org/10.1146/annurev.immunol.021908.132710>
169. Kunkl A, Mortara L, Valle MT et al. Recognition of antigenic clusters of *Candida albicans* by T lymphocytes from human immunodeficiency virus-infected persons. *J Infect Dis* 178:488-496, 1998. DOI: <http://dx.doi.org/10.1086/515620>
170. Lasker BA, Elie CM, Lott TJ et al. Molecular epidemiology of *Candida albicans* strains isolated from the oropharynx of HIV-positive patients at successive clinic visits. *Med Mycol* 39:341-352, 2001. PMID: 11556764 DOI: <http://dx.doi.org/10.1080/714031035>
171. Launay O, Lortholary O, Bouges-Michel C, Jarrousse B, Bentata M, Guillemin L. Candidemia: a nosocomial complication in adults with late-stage AIDS. *Clin Infect Dis* 26:1134-1141, 1998. PMID: 9597242
172. Ledergerber B, Egger M, Telenti A. AIDS-related opportunistic illness and potent antiretroviral therapy. *JAMA* 283:2653-2654, 2000. PMID: 10819936
173. LeibundGut-Landmann S, Grob O, Robinson J et al. Syk- and CARD-9-dependent coupling of innate immunity to the induction of T helper cells that produce interleukin 17. *Nat Immunol* 8:630-638, 2007. DOI: <http://dx.doi.org/10.1038/ni1460>
174. Leigh JE, Barousse M, Swoboda RK et al. *Candida*-specific systemic cell-mediated immune reactivities in human immunodeficiency virus-positive persons with mucosal candidiasis. *J Infect Dis* 183:277-285, 2001. PMID: 11120933 DOI: <http://dx.doi.org/10.1086/317944>
175. Leigh JE, Steele C, Wormley FL, Jr. et al. Th1/Th2 cytokine expression in saliva of HIV-positive and HIV-negative individuals: a pilot study in HIV-positive individuals with oropharyngeal candidiasis. *J Acquir Immune Defic Syndr Hum Retrovirol* 19:373-380, 1998.
176. Lenander-Lumikari M and Johansson I. Effect of saliva composition on growth of *Candida albicans* and *Torulopsis glabata*. *Oral Microbiol Immunol* 10:233-240, 1995. DOI: <http://dx.doi.org/10.1111/j.1399-302X.1995.tb00148.x>
177. Levitz SM. Interactions of toll-like receptors with fungi. *Microbes Infect* 6:1351-1355, 2004. PMID: 15596119 DOI: <http://dx.doi.org/10.1016/j.micinf.2004.08.014>
178. Levitz SM, Mathews HL, Murphy JW. Direct antimicrobial activity of T cells. *Immunity Today* 16:387-391, 1995. PMID: 7546195 DOI: [http://dx.doi.org/10.1016/0167-5699\(95\)80007-7](http://dx.doi.org/10.1016/0167-5699(95)80007-7)
179. Li J, Farthing PM, Thornhill MH. Cytokine regulation of major histocompatibility complex antigen expression by human oral and skin keratinocytes. *Arch Oral Biol* 41:533-538, 1996. PMID: 8937643 DOI: [http://dx.doi.org/10.1016/0003-9969\(96\)00026-X](http://dx.doi.org/10.1016/0003-9969(96)00026-X)
180. Liew FY, Xu D, Brint EK. Negative regulation of Toll-like receptor-mediated immune responses. *Nat Rev Immunol* 5:446-458, 2005. PMID: 15928677 DOI: <http://dx.doi.org/10.1038/nri1630>
181. Lifson AR, Hilton JF, Westenhoe JL et al. Time from HIV seroconversion to oral candidiasis or hairy leukoplakia among homosexual and bisexual men enrolled in three prospective cohorts. *AIDS* 8:73-79, 1994. PMID: 8011239 DOI: <http://dx.doi.org/10.1097/00002030-199401000-00011>
182. Lilly EA, Hart DJ, Leigh JE et al. Tissue-associated cytokine expression in HIV-positive persons with oropharyngeal candidiasis. *J Infect Dis* 190:605-612, 2004. PMID: 15243938 DOI: <http://dx.doi.org/10.1086/422154>
183. Lin AL, Johnson DA, Patterson TF et al. Salivary anticandidal activity and saliva composition in an HIV-infected cohort. *Oral Microbiol Immunol* 16:270-278, 2001. DOI: <http://dx.doi.org/10.1034/j.1399-302x.2001.016005270.x>
184. Liu YJ, Kanzler H, Soumelis V, Gilliet M. Dendritic cell lineage, plasticity and cross-regulation. *Nature Immunol* 2:585-589, 2001. DOI: <http://dx.doi.org/10.1038/89726>
185. Lombardi T, Hauser C, Budtz-Jørgensen E. Langerhans cells: structure, function and role in oral pathological conditions. *J Oral Pathol Med* 22:193-202, 1993. PMID: 8315598
186. Lopez-Dupla M, Mora Sanz P, Pintado G et al. Clinical, endoscopic, immunologic, and therapeutic aspects of oropharyngeal and esophageal candidiasis in HIV-infected patients: a survey of 114 cases. *Am J Gastroenterol* 87:1771-1776, 1992.
187. Lu L, Hsieh M, Oriss TB et al. Generation of DC from mouse spleen cell cultures in response to GM-CSF: immunophenotypic and functional analysis. *Immunology* 84:127-134, 1995.
188. Lundqvist C, Baranov V, Teglund S, Hammarström S, Hammarström ML. Cytokine profile and ultrastructure of intraepithelial gamma delta T cells in chronically inflamed human gingiva suggest a cytotoxic effector function. *J Immunol* 153:2302-2312, 1994. PMID: 8051426
189. Lupetti A, Guzzi G, Paladini A, Swart K, Campa M, Senesi S. Molecular typing of *Candida albicans* in oral candidiasis: karyotype epidemiology with human immunodeficiency virus-seropositive patients in comparison with that with healthy carriers. *J Clin Microbiol* 33:1238-1242, 1995. PMID: 7615734
190. Ma CS, Chew GYJ, Simpson N et al. Deficiency of Th17 cells in hyper IgE syndrome due to mutations in STAT3. *J Exp Med* 205:1551-1557, 2008. DOI: <http://dx.doi.org/10.1084/jem.20080218>

191. Macatonia SE, Lau R, Patterson S, Pinching AJ, Knight SC. Dendritic cell infection, depletion and dysfunction in HIV-infected individuals. *Immunology* 71:38-45, 1990. PMID: 2145214
192. MacDonald AS and Pearce EJ. Cutting edge: polarized Th cell response induction by transferred antigen-pulsed dendritic cells is dependent on IL-4 or IL-12 production by recipient cells. *J Immunol* 168:3127-3130, 2002. PMID: 11907061
193. Maenza JR, Merz WG, Romagnoli MJ, Keruly JC, Moore RD, Gallant JE. Infection due to fluconazole-resistant *Candida* in patients with AIDS: prevalence and microbiology. *Clin Infect Dis* 24:28-34, 1997. PMID: 8994752
194. Makala LH and Nagasawa H. Dendritic cells: a specialized complex system of antigen presenting cells. *J Vet Med Sci* 64:181-193, 2002. PMID: 11999435
195. Mandel ID, Barr CE, Turgeon L. Longitudinal study of parotid saliva in HIV-1 infection. *J Oral Pathol Med* 21:209-213, 1992. PMID: 1403836 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1992.tb00103.x>
196. Marcotte H and Lavoie MC. Oral microbial ecology and the role of salivary immunoglobulin A. *Microbiol Mol Biol Rev* 62:71-109, 1998. PMID: 9529888
197. Margiotta V, Campisi G, Mancuso S, Accurso V, Abbadessa V. HIV infection: oral lesions, CD4+ cell count and viral load in an Italian study population. *J Oral Pathol Med* 28:173-177, 1999.
198. Martinez FO, Helming L, Gordon S. Alternative activation of macrophages: an immunologic functional perspective. *Ann Rev Immunol* 27:451-483, 2009.
199. Martins MD, Lozano-Chiu M, Rex JH. Declining rates of oropharyngeal candidiasis and carriage of *Candida albicans* associated with trends toward reduced rates of carriage of fluconazole-resistant *C. albicans* in human immunodeficiency virus-infected patients. *Clin Infect Dis* 27:1291-1294, 1998. PMID: 9827284 DOI: <http://dx.doi.org/10.1086/515006>
200. McCarthy GM. Host factors associated with HIV-related oral candidiasis. A review. *Oral Surg Oral Med Oral Pathol* 73:181-186, 1992. PMID: 1532236 DOI: [http://dx.doi.org/10.1016/0030-4220\(92\)90192-S](http://dx.doi.org/10.1016/0030-4220(92)90192-S)
201. McCarthy GM, Mackie ID, Koval J, Sandhu HS, Daley TD. Factors associated with increased frequency of HIV-related oral candidiasis. *J Oral Pathol Med* 20:332-336, 1991. PMID: 1680189 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1991.tb00940.x>
202. McGuirk P and Mills KHG. Pathogen-specific regulatory T cells provoke a shift in the Th1/Th2 paradigm in immunity to infectious diseases. *Trends Immunol* 23:450-455, 2002. DOI: [http://dx.doi.org/10.1016/S1471-4906\(02\)02288-3](http://dx.doi.org/10.1016/S1471-4906(02)02288-3)
203. McIlroy D, Autran B, Clauvel JP, Oksenhendler E, Debré P, Hosmalin A. Low CD83, but normal MHC class II and costimulatory molecule expression, on spleen dendritic cells from HIV+ patients. *AIDS Res Hum Retroviruses* 14:505-513, 1998. PMID: 9566553
204. Medzhitov R. Recognition of microorganisms and activation of the immune response. *Nature* 449:819-826, 2007. PMID: 17943118 DOI: <http://dx.doi.org/10.1038/nature06246>
205. Medzhitov R and Janeway CA, Jr. Advances in Immunology: innate immunity. *New Engl J Med* 343:338-344, 2000.
206. Meiller TF, Jabra-Rizk MA, Baqui A et al. Oral *Candida dubliniensis* as a clinically important specie in HIV-seropositive patients in the United States. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 88:573-580, 1999. DOI: [http://dx.doi.org/10.1016/S1079-2104\(99\)70088-0](http://dx.doi.org/10.1016/S1079-2104(99)70088-0)
207. Melo NR, Taguchi H, Culhari VP et al. Oral candidiasis of HIV-infected children undergoing sequential HIV therapies. *Med Mycol* 47:149-156, 2009. PMID: 18651304 DOI: <http://dx.doi.org/10.1080/13693780802195315>
208. Mencacci A, Del Sero G, Cenci E et al. Endogenous interleukin 4 is required for development of protective CD4+ T helper type 1 cell responses to *Candida albicans*. *J Exp Med* 187:307-317, 1998. PMID: 9449711 DOI: <http://dx.doi.org/10.1084/jem.187.3.307>
209. Mezzaroma I, Carlesimo M, Pinter E et al. Long-term evaluation of T-cell subsets and T-cell function after HAART in advanced stage HIV-1 disease. *AIDS* 13:1187-1193, 1999. PMID: 10416521 DOI: <http://dx.doi.org/10.1097/00002030-199907090-00006>
210. Mills KHG. Regulatory T cells: friend or foe in immunity to infection? *Nat Rev Immunol* 4:841-855, 2004. PMID: 15516964 DOI: <http://dx.doi.org/10.1038/nri1485>
211. Miziara ID and Weber R. Oral lesions as predictors of highly active antiretroviral therapy failure in Brazilian HIV-infected children. *J Oral Pathol Med* 37:99-106, 2008. DOI: <http://dx.doi.org/10.1111/j.1600-0714.2007.00598.x>
212. Mohamed AM. Ultrastructural aspects of chronic oral candidosis. *J Oral Pathol* 4:180-194, 1975. PMID: 811778 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1975.tb01741.x>
213. Monod M, Hube B, Hess D, Sanglard D. Differential regulation of SAP8 and SAP9, which encode two new members of the secreted aspartic proteinase family in *Candida albicans*. *Microbiology* 144:2731-2737, 1998. PMID: 9802014 DOI: <http://dx.doi.org/10.1099/00221287-144-10-2731>
214. Mostefaoui Y, Claveau I, Rouabhia M. In vitro analyses of tissue structure and interleukin-1beta expression and production by human oral mucosa in response to *Candida albicans* infections. *Cytokine* 25:162-171, 2004. PMID: 15162833 DOI: <http://dx.doi.org/10.1016/j.cyto.2003.11.015>
215. Müller F, Froland SS, Hvatum M, Radl J, Brandtzaeg P. Both IgA subclasses are reduced in parotid saliva from patients with AIDS. *Clin Exp Immunol* 83:203-209, 1991. PMID: 1899629
216. Munro CA and Hube B. Anti-fungal therapy at the HAART of viral therapy. *Trends Microbiol* 10:173-177, 2002. PMID: 11912023
217. Myers TA, Leigh JE, Arribas AR et al. Immunohistochemical evaluation of T cells in oral lesions from human immunodeficiency virus-positive persons with oropharyngeal candidiasis. *Infect Immun* 71:956-963, 2003. PMID: 12540578 DOI: <http://dx.doi.org/10.1128/IAI.71.2.956-963.2003>
218. Nakanishi K, Yoshimoto T, Tsutsui H, Okamura H. Interleukin-18 regulates both Th1 and Th2 responses. *Ann Rev Immunol* 19:423-474, 2001. DOI: <http://dx.doi.org/10.1146/annurev.immunol.19.1.423>
219. Netea MG, Gijzen L, Coolen N et al. Human dendritic cells are less potent at killing *Candida albicans* than both monocytes and macrophages. *Microbes Infect* 6:985-989, 2004. DOI: <http://dx.doi.org/10.1016/j.micinf.2004.05.013>
220. Netea MG, van der Graaf C, van der Meer JW, Kullberg BJ. Toll-like receptors and the host defense against microbial pathogens: bringing specificity to the innate-immune system. *J Leukoc Biol* 75:749-755, 2004. PMID: 15075354
221. Newman SL and Holly A. *Candida albicans* is phagocytosed, killed, and processed for antigen presentation by human dendritic cells. *Infect Immun* 69:6813-6822, 2001. PMID: 11598054 DOI: <http://dx.doi.org/10.1128/IAI.69.11.6813-6822.2001>

222. Nielsen H, Bentsen KD, Højtved L et al. Oral candidiasis and immune status of HIV-infected patients. *J Oral Pathol Med* 23:140-143, 1994. PMID: 7912732 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1994.tb01102.x>
223. Nikawa H, Samaranyake LP, Tenovuo J, Pang KM, Hamada T. The fungicidal effect of human lactoferrin on *Candida albicans* and *Candida krusei*. *Arch Oral Biol* 38:1057-1063, 1993. PMID: 8141667 DOI: [http://dx.doi.org/10.1016/0003-9969\(93\)90167-K](http://dx.doi.org/10.1016/0003-9969(93)90167-K)
224. Nilsson J, Boasso A, Velilla PA et al. HIV-1-driven regulatory T-cell accumulation in lymphoid tissues is associated with disease progression in HIV/AIDS. *Blood* 108:3808-3817, 2006. PMID: 16902147 DOI: <http://dx.doi.org/10.1182/blood-2006-05-021576>
225. Nottet HS, de Graaf L, de Vos NM et al. Phagocytic function of monocyte-derived macrophages is not affected by human immunodeficiency virus type 1 infection. *J Infect Dis* 168:84-91, 1993. DOI: <http://dx.doi.org/10.1093/infdis/168.1.84>
226. O'Brien WA. Resistance against reverse transcriptase inhibitors. *Clin Infect Dis* 30S:185-192, 2000.
227. Odden K, Schenck K, Hurlen B. High numbers of T cells in gingiva from patients with human immunodeficiency virus (HIV) infection. *J Oral Pathol Med* 24:413-419, 1995. PMID: 8537915 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1995.tb01211.x>
228. Ollert MW, Wende C, Görlich M et al. Increased expression of *Candida albicans* secretory proteinase, a putative virulence factor, in isolates from human immunodeficiency virus-positive patients. *J Clin Microbiol* 33:2543-2549, 1995. PMID: 8567880
229. Ortega KL, Ceballos-Salobreña A, Gaitán-Cepeda LA, Magalhães MG. Oral manifestations after immune reconstitution in HIV patients on HAART. *Int J STD AIDS* 19:305-308, 2008. PMID: 18482959 DOI: <http://dx.doi.org/10.1258/ijsa.2007.007261>
230. Pacanowsky J, Kahi S, Baillet M et al. Reduced blood CD123+ (lymphoid) and CD11c+ (myeloid) dendritic cell numbers in primary HIV-1 infection. *Blood* 98:3016-3021, 2001.
231. Palella FJ Jr., Delaney KM, Moorman AC et al. Declining morbidity and mortality among patients with advanced human immunodeficiency virus infection. HIV Outpatient Study Investigators. *N Engl J Med* 338:853-860, 1998. DOI: <http://dx.doi.org/10.1056/NEJM199803263381301>
232. Pakker NG, Roos MT, van Leeuwen R et al. Patterns of T-cell repopulation, virus load reduction, and restoration of T-cell function in HIV-infected persons during therapy with different antiretroviral agents. *J Acquir Immune Defic Syndr Hum Retrovirol* 16:318-326, 1997. PMID: 9420308 DOI: <http://dx.doi.org/10.1097/00042560-199712150-00002>
233. Palmer GD, Robinson PG, Challacombe SJ et al. Aetiological factors for oral manifestations of HIV. *Oral Dis* 2:193-197, 1996. PMID: 9081758 DOI: <http://dx.doi.org/10.1111/j.1601-0825.1996.tb00223.x>
234. Patterson S, English NR, Longhurst H et al. Analysis of human immunodeficiency virus type 1 (HIV-1) variants and levels of infection in dendritic and T cells from symptomatic HIV-1 infected patients. *J Gen Virol* 79:247-257, 1998. PMID: 9472609
235. Patton LL. Sensitivity, specificity, and positive predictive value of oral opportunistic infections in adults with HIV/AIDS as markers of immune suppression and viral burden. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 90:182-188, 2000. PMID: 10936837 DOI: <http://dx.doi.org/10.1067/moe.2000.108799>
236. Patton LL, McKaig RG, Strauss RP, Eron JJ, Jr. Oral manifestations of HIV in a southeast USA population. *Oral Dis* 4:164-169, 1998. DOI: <http://dx.doi.org/10.1111/j.1601-0825.1998.tb00274.x>
237. Patton LL, McKaig R, Strauss R, Rogers D, Eron JJ, Jr. Changing prevalence of oral manifestations of human immunodeficiency virus in the era of protease inhibitor therapy. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 89:299-304, 2000. DOI: [http://dx.doi.org/10.1016/S1079-2104\(00\)70092-8](http://dx.doi.org/10.1016/S1079-2104(00)70092-8)
238. Peiser M, Grützkau A, Wanner R, Kolde G. CD1a and CD1c cell sorting yields a homogeneous population of immature human Langerhans cells. *J Immunol Methods* 279:41-53, 2003. PMID: 12969546 DOI: [http://dx.doi.org/10.1016/S0022-1759\(03\)00257-6](http://dx.doi.org/10.1016/S0022-1759(03)00257-6)
239. Peña JM, Martínez-López MA, Arnalich F, Barbado FJ, Vázquez JJ. Esophageal candidiasis associated with acute infection due to human immunodeficiency virus: case report and review. *Rev Infect Dis* 13:872-875, 1991. PMID: 1962101
240. Petrosillo N, Viale P, Nicastrì E et al. Nosocomial bloodstream infections among human immunodeficiency virus-infected patients: incidence and risk factors. *Clin Infect Dis* 34:677-685, 2002. PMID: 11823956
241. Pillay D, Taylor S, Richman DD. Incidence and impact of resistance against approved antiretroviral drugs. *Rev Med Virol* 10:231-253, 2000. PMID: 10891871 DOI: [http://dx.doi.org/10.1002/1099-1654\(200007/08\)10:4<231::AID-RMV290>3.0.CO;2-P](http://dx.doi.org/10.1002/1099-1654(200007/08)10:4<231::AID-RMV290>3.0.CO;2-P)
242. Polyak S, Chen H, Hirsch D, George I, Hershberg R, Sperber K. Impaired class II expression and antigen uptake in monocytes after HIV-1 infection. *J Immunol* 159:2177-2188, 1997. PMID: 9278305
243. Pomarico L, Cerqueira DF, Soares RMA et al. Associations among the use of highly active antiretroviral therapy, oral candidiasis, oral *Candida* species and salivary immunoglobulin A in HIV-infected children. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 108:203-210, 2009. DOI: <http://dx.doi.org/10.1016/j.tripleo.2009.05.008>
244. Pope M. Mucosal dendritic cells and immunodeficiency viruses. *J Infect Dis* 179:S427-430, 1999. DOI: <http://dx.doi.org/10.1086/314798>
245. Pope M, Elmore D, Ho D, Marx P. Dendritic cell-T cell mixtures, isolated from the skin and mucosae of macaques, support the replication of SIV. *AIDS Res Hum Retroviruses* 13:819-827, 1997. PMID: 9197376 DOI: <http://dx.doi.org/10.1089/aid.1997.13.819>
246. Powderly WG, Robinson K, Keath EJ. Molecular epidemiology of recurrent oral candidiasis in human immunodeficiency virus-positive patients: evidence for two patterns of recurrence. *J Infect Dis* 168:463-466, 1993. PMID: 8335986
247. Pulendran B, Kumar P, Cutler CW, Mohamadzadeh M, Van Dyke T, Banchereau J. Lipopolysaccharides from distinct pathogens induce different classes of immune responses in vivo. *J Immunol* 167:5067-5076, 2001.
248. Pulendran B, Smith JL, Caspary G et al. Distinct dendritic cell subsets differentially regulate the class of immune response in vivo. *Proc Natl Acad Sci USA* 96:1036-1041, 1999. DOI: <http://dx.doi.org/10.1073/pnas.96.3.1036>
249. Qureshi MN, Barr CE, Seshamma T, Reidy J, Pomerantz RJ, Bagasra O. Infection of oral mucosal cells by human immunodeficiency virus type 1 in seropositive persons. *J Infect Dis* 171:190-193, 1995. DOI: <http://dx.doi.org/10.1093/infdis/171.1.190>

250. Rabeneck L and Laine L. Esophageal candidiasis in patients infected with the immunodeficiency virus. A decision analysis to assess cost-effectiveness of alternative management strategies. *Arch Intern Med* 154:2705-2710, 1994. DOI: <http://dx.doi.org/10.1001/archinte.1994.00420230096011>
251. Radeke HH, von Wenckstern H, Stoldtner K, Sauer B, Hammer S, Kleuser B. Overlapping signaling pathways of sphingosine 1-phosphate and TGF-beta in the murine Langerhans cell line XS52. *J Immunol* 174:2778-2786, 2005. PMID: 15728487
252. Ramírez-Amador V, Esquivel-Pedraza L, Sierra-Madero J, Anaya-Saavedra G, González-Ramírez I, Ponce-de-León S. The changing clinical spectrum of human immunodeficiency virus (HIV)-related oral lesions in 1,000 consecutive patients: A 12-year study in a referral center in Mexico. *Medicine (Baltimore)* 82:39-50, 2003.
253. Ramírez-Amador V, Ponce-de-León S, Anaya-Saavedra G, Crabtree Ramírez B, Sierra-Madero J. Oral lesions as clinical markers of highly active antiretroviral therapy failure: a nested case-control study in Mexico City. *Clin Infect Dis* 45:925-932, 2007.
254. Rayhan R, Xu L, Santarpia RP 3rd, Tylanda CA, Pollock JJ. Antifungal activities of salivary histidine-rich polypeptides against *Candida albicans* and other oral yeast isolates. *Oral Microbiol Immunol* 7:51-52, 1992. PMID: 1528625 DOI: <http://dx.doi.org/10.1111/j.1399-302X.1992.tb00020.x>
255. Redding SW, Pfaller MA, Messer SA et al. Variations in fluconazole susceptibility and DNA subtyping of multiple *Candida albicans* colonies from patients with AIDS and oral candidiasis suffering one or more episodes of infection. *J Clin Microbiol* 35:1761-1765, 1997. PMID: 9196188
256. Reef SE and Mayer KH. Opportunistic candidal infections in patients infected with human immunodeficiency virus: prevention issues and priorities. *Clin Infect Dis* 21S:99-102, 1995.
257. Reibel J, Dabelsteen E, Kenrad B, Buschard K. Pattern of distribution of T lymphocytes, Langerhans cells and HLA-DR bearing cells in normal human oral mucosa. *Scand J Dent Res* 93:513-521, 1985. PMID: 2937133 DOI: <http://dx.doi.org/10.1111/j.1600-0722.1985.tb01349.x>
258. Reichart PA, Samaranyake LP, Philipsen HP. Pathology and clinical correlates in oral candidiasis and its variants: a review. *Oral Dis* 6:85-91, 2000. PMID: 10702784
259. Reichart PA, Philipsen HP, Schmidt-Westhausen A, Samaranyake LP. Pseudomembranous oral candidiasis in HIV infection: ultrastructural findings. *J Oral Pathol Med* 24:276-281, 1995. PMID: 7562665
260. Reichart PA, Schmidt-Westhausen A, Samaranyake LP, Philipsen HP. *Candida*-associated palatal papillary hyperplasia in HIV infection. *J Oral Pathol Med* 23:403-405, 1994. PMID: 7823300 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1994.tb00085.x>
261. Reichart PA, Weigel D, Schmidt-Westhausen A, Pohle HD. Exfoliative cheilitis (EC) in AIDS: association with *Candida* infection. *J Oral Pathol Med* 26:290-293, 1997. PMID: 9234190
262. Reid DM, Gow NA, Brown GD. Pattern recognition: recent insights from Dectin-1. *Curr Opin Immunol* 21:30-37, 2009.
263. Rhodus NL, Bloomquist C, Liljemark W, Bereuter J. Prevalence, density, and manifestations of oral *Candida albicans* in patients with Sjögren's syndrome. *J Otolaryngol* 26:300-305, 1997.
264. Ripeau JS, Fiorillo M, Aumont F, Belhumeur P, de Repentigny L. Evidence for differential expression of *Candida albicans* virulence genes during oral infection in intact and human immunodeficiency virus type 1-transgenic mice. *J Infect Dis* 185:1094-1102, 2002. PMID: 11930319 DOI: <http://dx.doi.org/10.1086/340035>
265. Rissoan MC, Soumelis V, Kadowaki N et al. Reciprocal control of T helper cell and dendritic cell differentiation. *Science* 283:1124-1125, 1999. DOI: <http://dx.doi.org/10.1126/science.283.5405.1183>
266. Robinson PG, Challacombe SJ, Sheiham A, Zakrzewska JM. Is erythematous candidiasis associated with advanced HIV disease? *Oral Dis* 3S:116-118, 1997. DOI: <http://dx.doi.org/10.1111/j.1601-0825.1997.tb00339.x>
267. Robinson MJ, Osorio F, Rosas M et al. Dectin-2 is a Syk-coupled pattern recognition receptor crucial for Th17 responses to fungal infection. *J Exp Med* 206:2037-2051, 2009. DOI: <http://dx.doi.org/10.1084/jem.20082818>
268. Romagnoli P, Pimpinelli N, Mori M, Reichart PA, Eversole LR, Ficarra G. Immunocompetent cells in oral candidiasis of HIV-infected patients: an immunohistochemical and electron microscopical study. *Oral Dis* 3:99-105, 1997. PMID: 9467350
269. Romani N, Gruner S, Brang D et al. Proliferating dendritic cell progenitors in human blood. *J Exp Med* 180:83-93, 1994. DOI: <http://dx.doi.org/10.1084/jem.180.1.83>
270. Royce RA, Luckmann RS, Fusaro RE, Winkelstein W, Jr. The natural history of HIV-1 infection: staging classifications of disease. *AIDS* 5:355-364, 1991. PMID: 1676278
271. Rutherford GW, Sangani PR, Kennedy GE. Three- or four- versus two-drug antiretroviral maintenance regimens for HIV infection. *Cochrane Database Syst Rev* 4:CD002037, 2003. PMID: 14583945
272. Samaranyake LP and Holmstrup P. Oral candidiasis and human immunodeficiency virus infection. *J Oral Pathol Med* 18:554-564, 1989. PMID: 2695620 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1989.tb01552.x>
273. Samaranyake LP, Hughes A, Weetman DA, MacFarlane TW. Growth and acid production of *Candida* species in human saliva supplemented with glucose. *J Oral Pathol* 15:251-254, 1986. PMID: 3091791 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1986.tb00617.x>
274. Samaranyake YH, Samaranyake LP, Pow EH, Beena VT, Yeung KW. Antifungal effects of lysozyme and lactoferrin against genetically similar, sequential *Candida albicans* isolates from a human immunodeficiency virus-infected southern Chinese cohort. *J Clin Microbiol* 39:3296-3302, 2001. PMID: 11526166 DOI: <http://dx.doi.org/10.1128/JCM.39.9.3296-3302.2001>
275. Sangeorzan JA, Bradley SF, He X et al. Epidemiology of oral candidiasis in HIV-infected patients: colonization, infection, treatment, and emergence of fluconazole resistance. *Am J Med* 97:339-346, 1994. PMID: 7942935
276. Sansonetti PJ. The innate signaling of dangers and the dangers of innate signaling. *Nat Immunol* 7:1237-1242, 2006. PMID: 17110939 DOI: <http://dx.doi.org/10.1038/ni1420>
277. Schaller M, Mailhammer R, Korting HC. Cytokine expression induced by *Candida albicans* in a model of cutaneous candidosis based on reconstituted human epidermis. *J Med Microbiol* 51:672-676, 2002.
278. Schmidt-Westhausen AM, Priepeke F, Bergmann FJ, Reichart PA. Decline in the rate of oral opportunistic infections following introduction of highly active antiretroviral therapy. *J Oral Pathol Med* 29:336-341, 2000. PMID: 10947250 DOI: <http://dx.doi.org/10.1034/j.1600-0714.2000.290708.x>
279. Schoenberger SP, Toes RE, van der Voort EI, Offringa R, Melief CJ. T-cell help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. *Nature* 393:480-483, 1998. PMID: 9624005 DOI: <http://dx.doi.org/10.1038/31002>

280. Schorling SR, Kortinga HC, Froschb M, Mühlshchel FA. The role of *Candida dubliniensis* in oral candidiasis in human immunodeficiency virus-infected individuals. *Crit Rev Microbiol* 26:59-68, 2000. DOI: <http://dx.doi.org/10.1080/10408410091154183>
281. Schuman P, Sobel JD, Ohmit SE et al. Mucosal candidal colonization and candidiasis in women with or at risk for human immunodeficiency virus infection. HIV Epidemiology Research Study (HERBS) Group. *Clin Infect Dis* 27:1161-1167, 1998. DOI: <http://dx.doi.org/10.1086/514979>
282. Segal AW. How neutrophils kill microbes. *Ann Rev Immunol* 23:197-223, 2005. DOI: <http://dx.doi.org/10.1146/annurev.immunol.23.021704.115653>
283. Séguier S, Godeau G, Brousse N. Immunohistological and morphometric analysis of intra-epithelial lymphocytes and Langerhans cells in healthy and diseased human gingival tissues. *Arch Oral Biol* 45:441-452, 2000. PMID: 10775673
284. Séguier S, Godeau G, Leborgne M, Pivert G, Brousse N. Quantitative morphological analysis of Langerhans cells in healthy and diseased human gingiva. *Arch Oral Biol* 45:1073-1081, 2000. PMID: 11084147
285. Selik RM, Starcher ET, Curran JW. Opportunistic diseases reported in AIDS patients: frequencies, associations, and trends. *AIDS* 1:175-182, 1987. PMID: 2831912
286. Sepkowitz KA. Effect of HAART on natural history of AIDS-related opportunistic disorders. *Lancet* 351:228-230, 1998. PMID: 9457088 DOI: [http://dx.doi.org/10.1016/S0140-6736\(05\)78279-9](http://dx.doi.org/10.1016/S0140-6736(05)78279-9)
287. Sherwood J, Gow NA, Gooday GW, Gregory DW, Marshall D. Contact sensing in *Candida albicans*: a possible aid to epithelial penetration. *J Med Vet Mycol* 30:461-469, 1992.
288. Shiboski CH. Epidemiology of HIV-related oral manifestations in women: a review. *Oral Dis* 3S:18-27, 1997.
289. Shortman K and Liu YJ. Mouse and human dendritic cell subtypes. *Nat Rev Immunol* 2:151-161, 2002. PMID: 11913066 DOI: <http://dx.doi.org/10.1038/nri746>
290. Soukka T, Tenovu J, Lenander-Lumikari M. Fungicidal effect of human lactoferrin against *Candida albicans*. *FEMS Microbiol Lett* 69:223-228, 1992. PMID: 1555756 DOI: <http://dx.doi.org/10.1111/j.1574-6968.1992.tb05156.x>
291. Steele C and Fidel PL, Jr. Cytokine and chemokine production by human oral and vaginal epithelial cells in response to *Candida albicans*. *Infect Immun* 70:577-583, 2002. DOI: <http://dx.doi.org/10.1128/IAI.70.2.577-583.2002>
292. Steele C, Leigh J, Swoboda R, Ozenci H, Fidel PL, Jr. Potential role for a carbohydrate moiety in anti-*Candida* activity of human oral epithelial cells. *Infect Immun* 69:7091-1099, 2001. DOI: <http://dx.doi.org/10.1128/IAI.69.11.7091-7099.2001>
293. Steidley KE, Thompson SH, McQuade MJ, Strong SL, Scheidt MJ, Van Dyke TE. A comparison of T4:T8 lymphocyte ratio in the periodontal lesion of healthy and HIV-positive patients. *J Periodontol* 63:753-756, 1992. PMID: 1361945
294. Stein DS, Korvick JA, Vermund SH. CD4+ lymphocyte cell enumeration for prediction of clinical course of human immunodeficiency virus disease: a review. *J Infect Dis* 165:352-363, 1992. PMID: 1346152
295. Steinhoff M, Brzoska T, Luger TA. Keratinocytes in epidermal immune responses. *Curr Opin Allergy Clin Immunol* 1:469-476, 2001. PMID: 11964729 DOI: <http://dx.doi.org/10.1097/00130832-200110000-00014>
296. Steinman RM, Adams JC, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. IV. Identification and distribution in mouse spleen. *J Exp Med* 141:804-820, 1975.
297. Steinman RM and Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. I. Morphology, quantitation, tissue distribution. *J Exp Med* 137:1142-1162, 1973. DOI: <http://dx.doi.org/10.1084/jem.137.5.1142>
298. Steinman RM and Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. II. Functional properties in vitro. *J Exp Med* 139:380-397, 1974. DOI: <http://dx.doi.org/10.1084/jem.139.2.380>
299. Steinman RM, Kaplan G, Witmer MD, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. V. Purification of spleen dendritic cells, new surface markers, and maintenance in vitro. *J Exp Med* 149:1-16, 1979. DOI: <http://dx.doi.org/10.1084/jem.149.1.1>
300. Steinman RM, Lustig DS, Cohn ZA. Identification of a novel cell type in peripheral lymphoid organs of mice. III. Functional properties in vivo. *J Exp Med* 139:1431-1445, 1974. DOI: <http://dx.doi.org/10.1084/jem.139.6.1431>
301. Stockinger B and Veldhoen M. Differentiation and function of the Th17 T cells. *Curr Opin Immunol* 19:281-286, 2007. PMID: 17433650
302. Strobl H, Scheinecker C, Riedl E et al. Identification of CD68+lin-peripheral blood cells with dendritic precursor characteristics. *J Immunol* 161:740-748, 1998. PMID: 9670950
303. Sullivan D, Haynes K, Bille J et al. Widespread geographic distribution of *Candida dubliniensis* strains in human immunodeficiency virus-infected individuals. *J Clin Microbiol* 35:960-964, 1997. PMID: 9157162
304. Suttmuller RP, Morgan ME, Netea MG, Grauer O, Adema GJ. Toll-like receptors on regulatory T cells: expanding immune regulation. *Trends Immunol* 27:387-393, 2006. PMID: 16814607
305. Sweet SP, Cookson S, Challacombe SJ. *Candida albicans* isolates from HIV-infected and AIDS patients exhibit enhanced adherence to epithelial cells. *J Med Microbiol* 43:452-457, 1995. PMID: 7473680 DOI: <http://dx.doi.org/10.1099/00222615-43-6-452>
306. Sweet SP, Rahman D, Challacombe SJ. IgA subclasses in HIV disease: dichotomy between raised levels in serum and decreased secretion rates in saliva. *Immunology* 86:556-559, 1995. PMID: 8567021
307. Tascini C, Baldelli F, Monari C et al. Inhibition of fungicidal activity of polymorphonuclear leukocytes from HIV-infected patients by interleukin (IL)-4 and IL-10. *AIDS* 10:477-483, 1996. PMID: 8724038
308. Tavittian A, Raufman JP, Rosenthal LE. Oral candidiasis as a marker for esophageal candidiasis in the acquired immunodeficiency syndrome. *Ann Intern Med* 104:54-55, 1986. PMID: 3940505 DOI: <http://dx.doi.org/10.7326/0003-4819-104-1-54>
309. Taylor PR, Martinez-Pomares L, Stacey M, Lin HH, Brown GD, Gordon S. Macrophage receptors and immune recognition. *Ann Rev Immunol* 23:901-944, 2005. DOI: <http://dx.doi.org/10.1146/annurev.immunol.23.021704.115816>
310. Telenti A and Rizzardi GP. Limits to potent antiretroviral therapy. *Rev Med Virol* 10: 385-393, 2000. DOI: [http://dx.doi.org/10.1002/1099-1654\(200011/12\)10:6<385::AID-RMV296>3.0.CO;2-1](http://dx.doi.org/10.1002/1099-1654(200011/12)10:6<385::AID-RMV296>3.0.CO;2-1)
311. Tenorio AR, Martinson J, Pollard D, Baum L, Landay A. The relationship of T-regulatory cell subsets to disease stage, immune activation, and pathogen-specific immunity in HIV infection. *J Acquir Immune Defic Syndr* 48:577-580, 2008. PMID: 18645514 DOI: <http://dx.doi.org/10.1097/QAI.0b013e31817bba5>

312. Tomee JF, Hiemstra PS, Heinzl-Wieland R, Kauffman HF. Antileukoprotease: an endogenous protein in the innate mucosal defense against fungi. *J Infect Dis* 176:740-747, 1997. PMID: 9291323
313. Trinchieri G and Sher A. Cooperation of Toll-like receptor signals in innate immune defence. *Nat Rev Immunol* 7:179-190, 2007. PMID: 17318230 DOI: <http://dx.doi.org/10.1038/nri2038>
314. Tsai H and Bobek LA. Human salivary histatins: promising anti-fungal therapeutic agents. *Crit Rev Oral Biol Med* 9:480-497, 1998. PMID: 9825223
315. Tumbarello M, Tacconelli E, de Gaetano Donati K, Morace G, Fadda G, Cauda R. Candidemia in HIV-infected subjects. *Eur J Clin Microbiol Infect Dis* 18:478-483, 1999. PMID: 10482024 DOI: <http://dx.doi.org/10.1007/s100960050327>
316. Tylenda CA, Larsen J, Yeh CK, Lane HC, Fox PC. High levels of oral yeasts in early HIV-1 infection. *J Oral Pathol Med* 18:520-524, 1989. PMID: 2575167 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1989.tb01355.x>
317. Umadevi KM, Ranganathan K, Pavithra S et al. Oral lesions among persons with HIV disease with and without highly active antiretroviral therapy in southern India. *J Oral Pathol Med* 36:136-141, 2007. PMID: 17305634 DOI: <http://dx.doi.org/10.1111/j.1600-0714.2006.00505.x>
318. Underhill DM and Ozinsky A. Toll-like receptors: key mediators of microbe detection. *Curr Opin Immunol* 14:103-110, 2002. PMID: 11790539
319. Vachot L, Williams VG, Bess JW Jr., Lifson JD, Robbiani M. Candida albicans-induced DC activation partially restricts HIV amplification in DCs and increases DC to T-cell spread of HIV. *J Acquir Immune Defic Syndr* 48:398-407, 2008. PMID: 18614931 DOI: <http://dx.doi.org/10.1097/QAI.0b013e3181776bc7>
320. van Dyke RB, Lee S, Johnson GM et al. Reported adherence as a determinant of response to highly active antiretroviral therapy in children who have human immunodeficiency virus infection. *Pediatrics* 109:e61, 2002. PMID: 11927734 DOI: <http://dx.doi.org/10.1542/peds.109.4.e61>
321. van Gisbergen K, Geijtenbeek T, van Kooyk Y. Close encounters of neutrophils and DCs. *Trends Immunol* 26:626-631, 2005. PMID: 16182604 DOI: <http://dx.doi.org/10.1016/j.it.2005.09.007>
322. van Kooyk Y and Geijtenbeek TB. DC-SIGN: escape mechanism for pathogens. *Nat Rev Immunol* 3:697-709, 2003. PMID: 12949494
323. van Loon LA, Krieg SR, Davidson CL, Bos JD. Quantification and distribution of lymphocyte subsets and Langerhans cells in normal human mucosa and skin. *J Oral Pathol Med* 18:197-201, 1989. PMID: 2570142 DOI: <http://dx.doi.org/10.1111/j.1600-0714.1989.tb00762.x>
324. van Vliet SJ, den Dunnen J, Gringhuis SI, Geijtenbeek TB, van Kooyk Y. Innate signaling and regulation of dendritic cell immunity. *Curr Opin Immunol* 19:435-440, 2007. PMID: 17629469 DOI: <http://dx.doi.org/10.1016/j.coi.2007.05.006>
325. van Wijngaerden E, de Saar V, de Graeve V et al. Nonadherence to highly active antiretroviral therapy: clinically relevant patient categorization based on electronic event monitoring. *AIDS Res Hum Retroviruses* 18:327-330, 2002. PMID: 11897033
326. Vargas KG and Joly S. Carriage frequency, intensity of carriage, and strains of oral yeast species vary in the progression to oral candidiasis in human immunodeficiency virus-positive individuals. *J Clin Microbiol* 40:341-350, 2002. PMID: 11825940 DOI: <http://dx.doi.org/10.1128/JCM.40.2.341-350.2002>
327. Vázquez-Torres A and Balish E. Macrophages in resistance to candidiasis. *Microbiol Mol Biol Rev* 61:170-192, 1997. PMID: 9184009
328. Villar CC, Kashleva H, Mitchell AP, Dongari-Bagtzoglou A. Invasive phenotype of *Candida albicans* affects the host proinflammatory response to infection. *Infect Immun* 73:4588-4595, 2005. PMID: 16040970 DOI: <http://dx.doi.org/10.1128/IAI.73.8.4588-4595.2005>
329. Volpe E, Servant N, Zollinger R et al. A critical function for transforming growth factor-beta, interleukin 23 and proinflammatory cytokines in driving and modulating human T(H)-17 responses. *Nat Immunol* 9:650-657, 2008.
330. Vudhichamnong K, Walker DM, Ryley HC. The effect of secretory immunoglobulin A on the in-vitro adherence of the yeast *Candida albicans* to human oral epithelial cells. *Arch Oral Biol* 27:617-621, 1982. PMID: 6753802 DOI: [http://dx.doi.org/10.1016/0003-9969\(82\)90184-4](http://dx.doi.org/10.1016/0003-9969(82)90184-4)
331. Vultaggio A, Lombardelli L, Giudizi MG et al. T cells specific for *Candida albicans* antigens and producing type 2 cytokines in lesional mucosa of untreated HIV-infected patients with pseudomembranous oropharyngeal candidiasis. *Microbes Infect* 10:166-174, 2008. PMID: 18249024 DOI: <http://dx.doi.org/10.1016/j.micinf.2007.11.004>
332. Wainberg MA and Clotet B. Review: immunologic response to protease inhibitor-based highly active antiretroviral therapy. *AIDS Patient Care STDS*. 21:609-620, 2007. PMID: 17919088
333. Weinberg ED. Iron and susceptibility to infectious disease. *Science* 184:952-956, 1974. PMID: 4596821 DOI: <http://dx.doi.org/10.1126/science.184.4140.952>
334. Weinberg A, Dickover R, Britto P et al. Continuous improvement in the immune system of HIV-infected children on prolonged antiretroviral therapy. *AIDS* 22:2267-2277, 2008. PMID: 18981766 DOI: <http://dx.doi.org/10.1097/QAD.0b013e3283189bb3>
335. Weinberg A, Krisanaprakornkit S, Dale BA. Epithelial antimicrobial peptides: review and significance for oral applications. *Crit Rev Oral Biol Med* 9:399-414, 1998. PMID: 9825219
336. Weindl G, Naglik JR, Kaesler S, Biedermaier T et al. Human epithelial cells establish direct antifungal defense through TLR4-mediated signaling. *J Clin Invest* 117:3664-3672, 2007. DOI: <http://dx.doi.org/10.1172/JCI28115>
337. Weinert M, Grimes RM, Lynch DP. Oral manifestations of HIV infection. *Ann Intern Med* 125:485-496, 1996. PMID: 8779462 DOI: <http://dx.doi.org/10.7326/0003-4819-125-6-199609150-00010>
338. Wenisch C, Parschalk B, Zedwitz-Liebenstein K, Graninger W, Rieger A. Dysregulation of the polymorphonuclear leucocyte-*Candida* spp. interaction in HIV-positive patients. *AIDS* 10:983-987, 1996. PMID: 8853731 DOI: <http://dx.doi.org/10.1097/00002030-199610090-00008>
339. Winning T, Gemmell E, Polak B, Savage NW, Seymour GJ, Walsh LJ. Expression of CD1a monocytes cultured with supernatants from periodontally diseased gingival epithelium cells. *Oral Dis* 2:247-252, 1996. PMID: 9171507 DOI: <http://dx.doi.org/10.1111/j.1601-0825.1996.tb00234.x>
340. Winzler C, Rovere P, Rescigno M et al. Maturation stages of mouse dendritic cells in growth factor-dependent long-term cultures. *J Exp Med* 185:317-328, 1997. PMID: 9016880 DOI: <http://dx.doi.org/10.1084/jem.185.2.317>

-
341. Witmer MD and Steinman RM. The anatomy of peripheral lymphoid organs with emphasis on accessory cells: light-microscopic immunocytochemical studies of mouse spleen, lymph node, and Peyer's patch. *Am J Anat* 170:465-481, 1984.
342. Wohlfert E and Belkaid Y. Role of endogenous and induced regulatory T cells during infections. *J Clin Immunol* 28:707-715, 2008. PMID: 18810611 DOI: <http://dx.doi.org/10.1007/s10875-008-9248-6>
343. Wu T, Samaranayake LP, Cao BY, Wang J. In vitro proteinase production by oral *Candida albicans* isolates from individuals with and without HIV infection and its attenuation by antimycotic agents. *J Med Microbiol* 44:311-316, 1996. PMID: 8606360 DOI: <http://dx.doi.org/10.1099/00222615-44-4-311>
344. Yeh CK, Dodds MW, Zuo P, Johnson DA. A population-based study of salivary lysozyme concentrations and candidal counts. *Arch Oral Biol* 42:25-31, 1997. PMID: 9134113 DOI: [http://dx.doi.org/10.1016/S0003-9969\(96\)00104-5](http://dx.doi.org/10.1016/S0003-9969(96)00104-5)
345. Yeh CK, Fox PC, Ship JA et al. Oral defense mechanisms are impaired early in HIV-1 infected patients. *J Acquir Immune Defic Syndr* 1:361-366, 1988. PMID: 3216316
346. Young JW and Steinman RM. Dendritic cells stimulate primary human cytolytic lymphocyte responses in the absence of CD4+ helper T cells. *J Exp Med* 171:1315-1332, 1990. PMID: 2139102 DOI: <http://dx.doi.org/10.1084/jem.171.4.1315>
347. Zhang N, Schröppel B, Lal G et al. Regulatory T cells sequentially migrate from inflamed tissues to draining lymph nodes to suppress the alloimmune response. *Immunity* 30:458-469, 2009. PMID: 19303390 DOI: <http://dx.doi.org/10.1016/j.immuni.2008.12.022>
348. Zuniga EI, McGavern DB, Pruneda-Paz JL, Teng C, Oldstone MB. Bone marrow plasmacytoid dendritic cells can differentiate into myeloid dendritic cells upon virus infection. *Nat Immunol* 5:1227-1234, 2004. DOI: <http://dx.doi.org/10.1038/ni1136>